

DILIsym® User Training - Gathering Mitochondrial Toxicity Data for DILIsym®

DILI-sim Team

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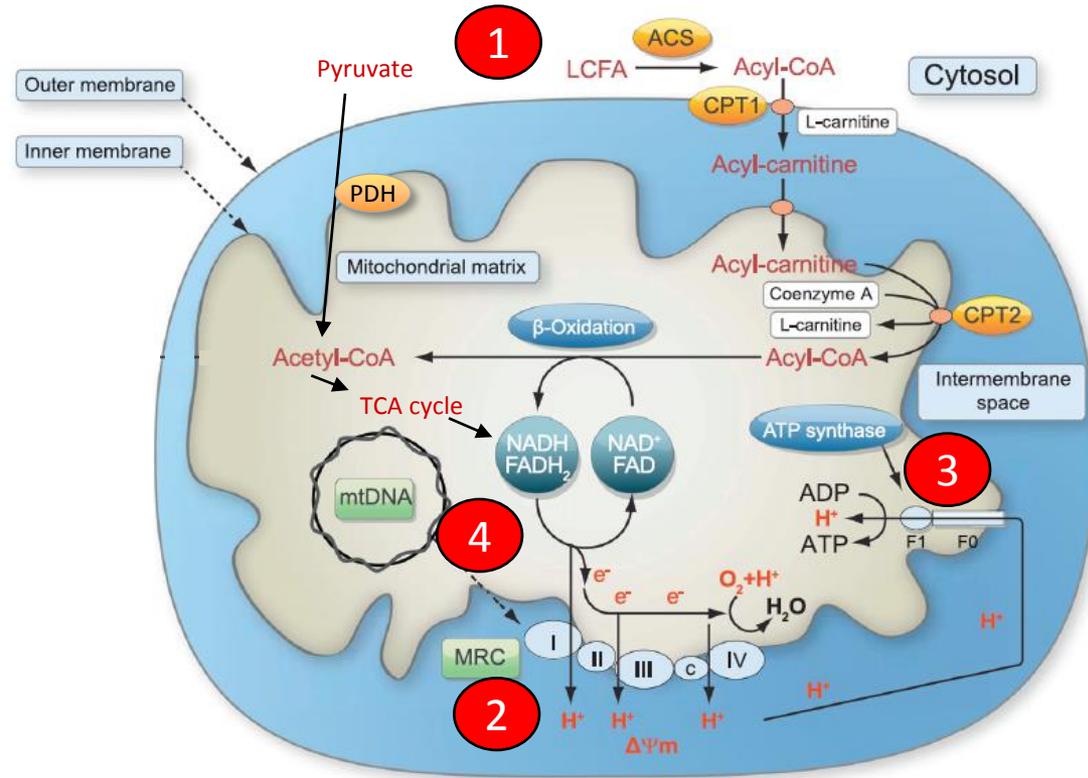
Goals for This Training Session

Participants should understand the following general concepts:

- The assay systems should be used to gather mitochondrial toxicity data for compounds to be represented in DILIsym®
- The conditions and general protocols for the mitochondrial toxicity assay used to provide data inputs for DILIsym®

Overview of Mitochondria Bioenergetics Biochemistry

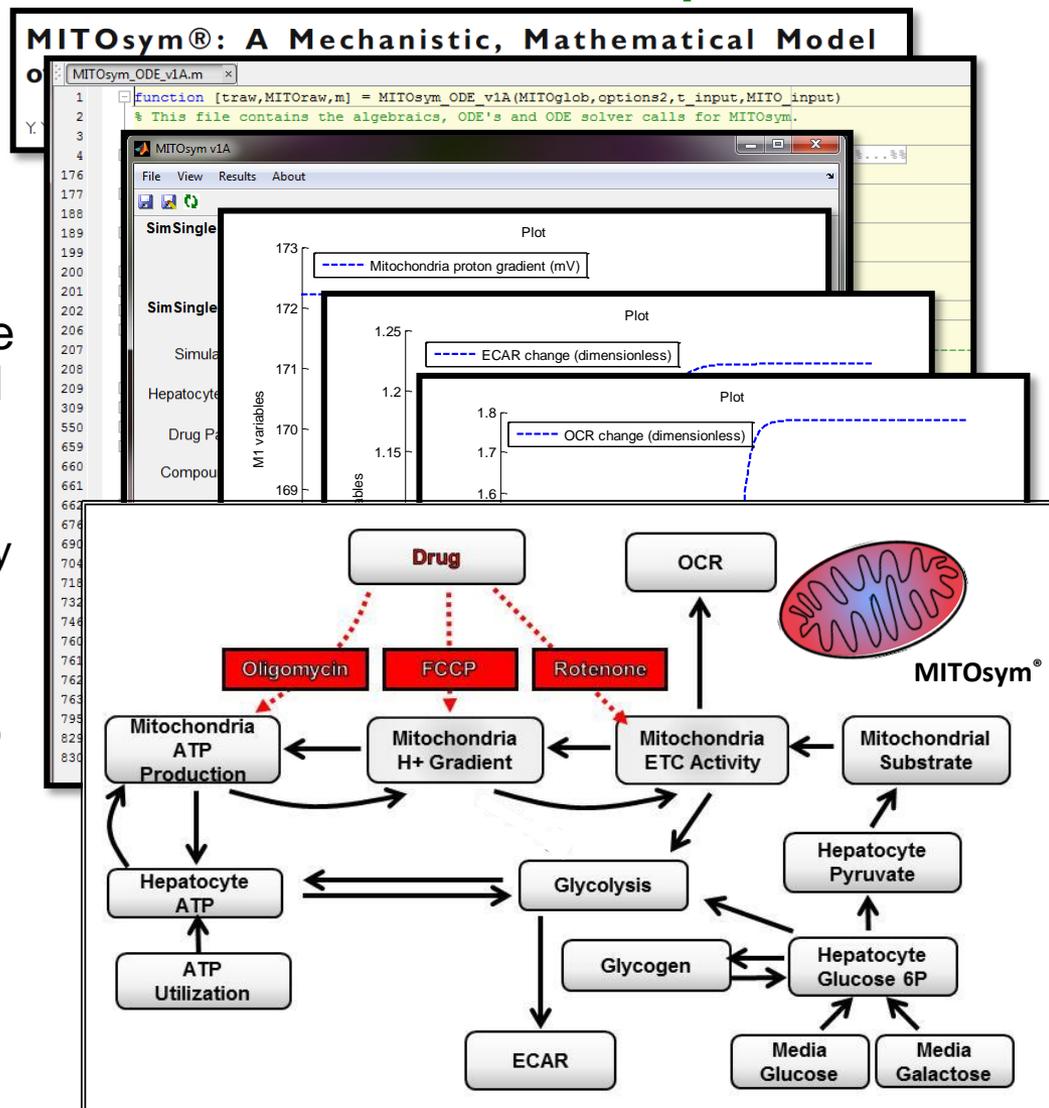
- 1 Metabolic substrate
 - Fatty acids via beta oxidation
 - Pyruvate via PDH
- 2 Electron Transport Chain (ETC)
 - Electrons donated from NADH and FADH₂
 - Protons pumped out of mitochondria, establishing H⁺ gradient ($\Delta\Psi_m$)
 - Electrons ultimately donated to O₂, accounting for energy-related respiration
- 3 ATP synthesis
 - H⁺ gradient used to drive ATP synthesis via ATP synthase
- 4 Mitochondrial DNA encodes multiple mitochondria proteins
 - Within ETC



Adapted from Begrich 2011

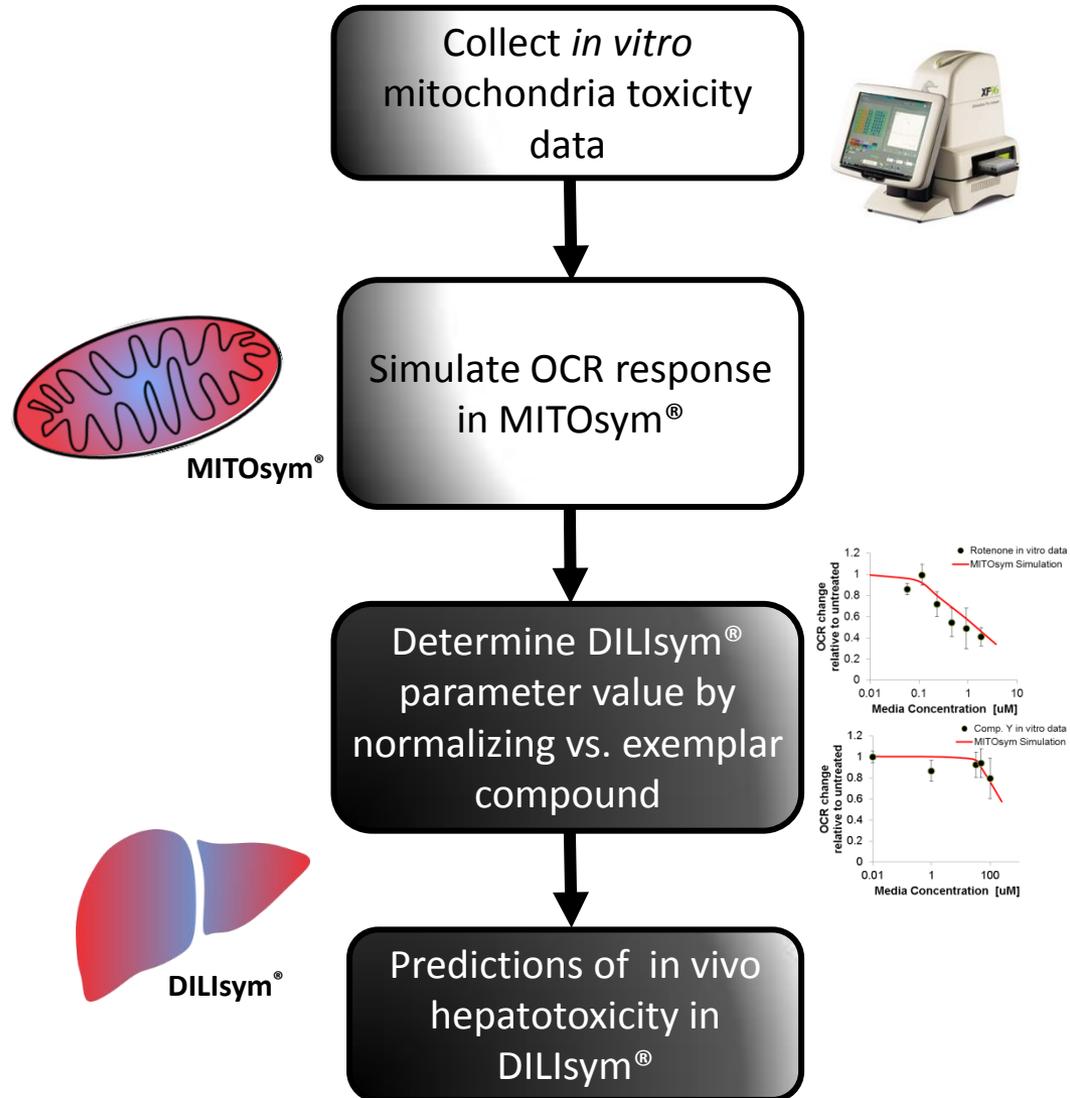
MITOsym[®] Is Designed to Support IVIVE DILI Predictions and Mechanistic Data Interpretation

- MITOsym[®] is a standalone model of hepatocyte bioenergetics
 - Yang et al. 2015
- MITOsym[®] can be used to facilitate predictions of hepatotoxicity based on *in vitro* cellular respiration data
 - Combine with DILIsym[®] model
 - Multiple cell types: HepG2, primary human hepatocytes, primary rat hepatocytes
- MITOsym[®] can be used to develop and explore hypotheses of the mechanisms underlying observed changes in respiration and glycolysis in hepatocytes
 - Comparison with exemplar drugs



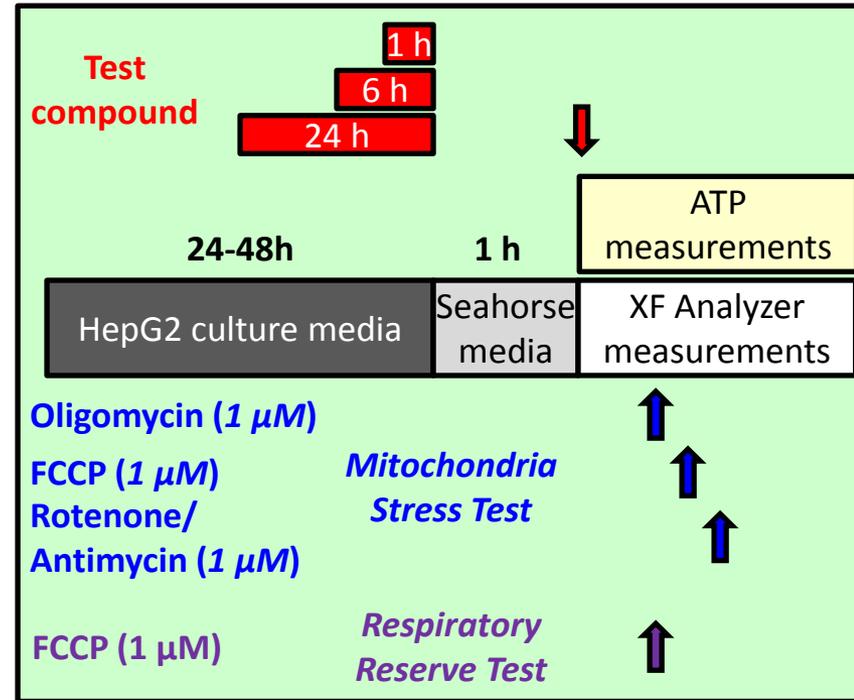
MITOsym[®] Developed to Support Identification of DILIsym[®] Toxicity Parameter Inputs

- Simulate Seahorse-based measurements of oxygen consumption rate (OCR) with MITOsym[®]
- Determine appropriate parameter values for mitochondria toxicity to be used in *in vivo* predictions with DILIsym[®]
- MITOsym[®] can be used to develop and explore hypotheses of the mechanisms underlying observed changes in respiration and glycolysis in hepatocytes
 - ETC inhibition, $\Delta\psi_m$ uncoupling, F1F0 ATPase inhibition, glycolysis inhibition



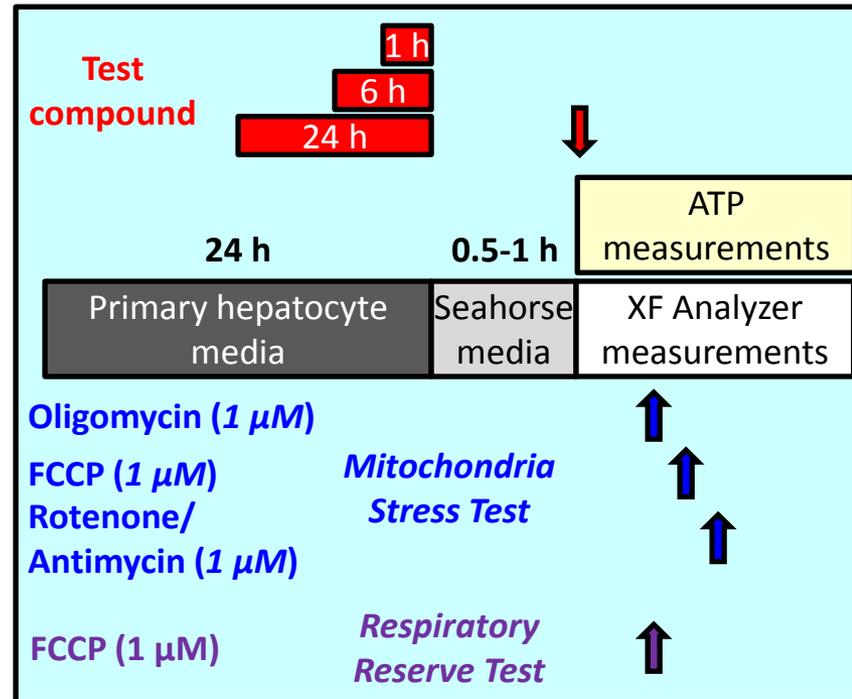
General XF Analyzer Protocol for DILIsym[®] Input Panel Mitochondria Measurements: HepG2

- HepG2 cells lack drug metabolism capacity
 - Exposure with parent, metabolites
 - Cells seeded into plates and cultured for 24-48 h
- Cells incubated in Seahorse media for 1 h
- Several possibilities for administering **test compound**
 - Direct injection, 1 h, 6 h, 24 h incubation
 - Depends on compound characteristics
 - Need to measure cell count with longer incubations
- **Mitochondria Stress Test** provides information on mitochondria toxicity mechanisms
- **Respiratory Reserve Test** can reveal modest ETC inhibition effects
- Measure ATP levels to validate OCR measures
- Measure intracellular compound concentrations coincident with OCR and ATP measures
 - Can estimate intracellular concentrations via PBPK methods



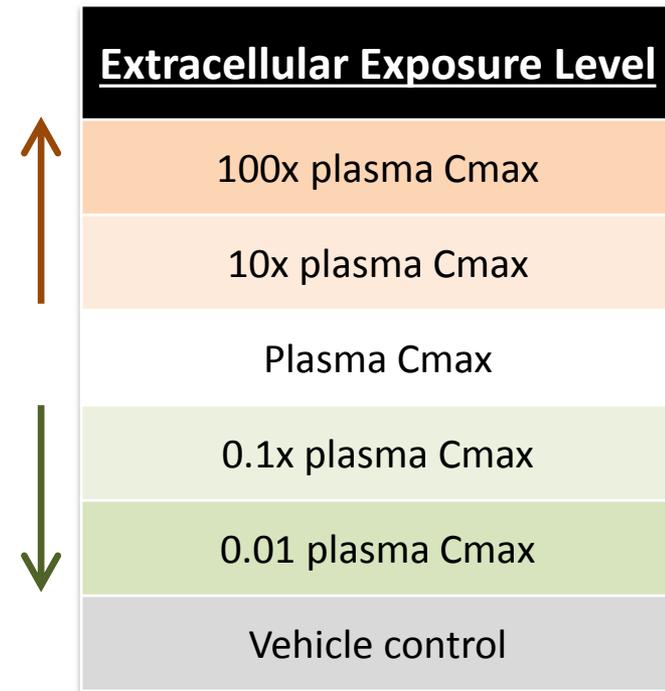
General XF Analyzer Protocol for DILIsym[®] Input Panel Mitochondria Measurements: Hepatocytes

- Primary hepatocytes are metabolically competent
 - Difficult to delineate effects of parent vs. metabolites
 - Culture condition requirements differ
- Cells incubated in Seahorse media for 0.5-1 h
- Several possibilities for administering **test compound**
 - Direct injection, 1 h, 6 h, 24 h incubation
 - Depends on compound characteristics
 - Need to measure cell count with longer incubations
- **Mitochondria Stress Test** provides information on mitochondria toxicity mechanisms
- **Respiratory Reserve Test** can reveal modest ETC inhibition effects
- Measure ATP levels to validate OCR measures
- Measure intracellular compound concentrations coincident with OCR and ATP measures
 - Active compound transporters can make it challenging to estimate intracellular concentrations via PBPK methods



The Recommended Extracellular Exposure Range is Based on Relevant Plasma Concentrations *in vivo*

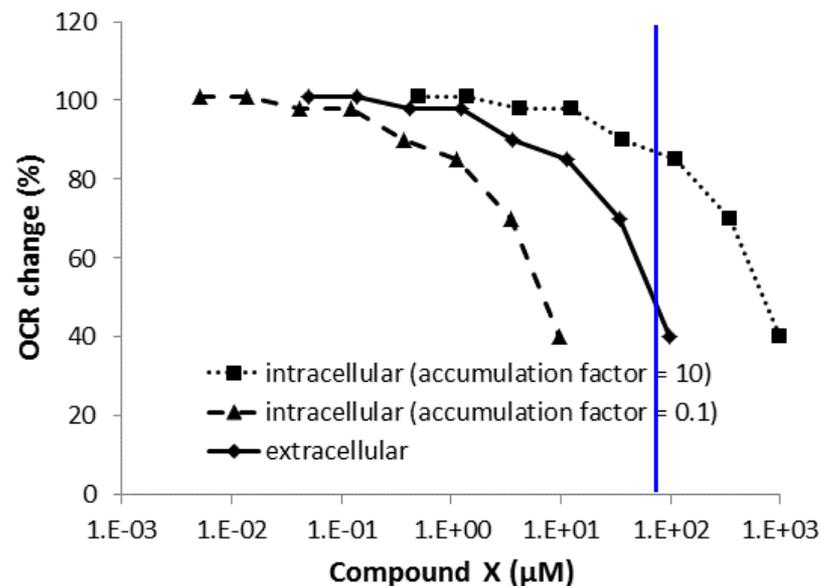
- Extracellular exposure ranges should be relative to the plasma concentrations, or predictions thereof
 - **This is critical to gathering useful data!**
 - *In vitro* studies are often done at extremely high concentrations
 - Contrary to the approach often taken, pushing the cells to a maximal response is not the best approach for this application
- Example table shown that blankets the predicted C_{max}
 - This example is applicable when clinical PK data are unavailable
- Range should be adjusted if more information is available
 - Clinical PK data can help set lower and upper bounds of plasma levels
 - High anticipated variability in exposure may warrant a broader range



Improved DILIsym[®] Parameter Values When Intracellular Compound Is Measured

- Most Seahorse oxygen consumption rate (OCR) or ROS/RNS data are expressed in an exposure-response relationship
 - OCR change on y-axis
 - Extracellular compound concentration on x-axis
- Numerous compounds have been shown to accumulate in liver
 - Potency relative to intracellular concentrations different than relative to extracellular
 - Intracellular \neq extracellular
- Basing parameter values on extracellular concentrations introduces inaccuracy for compounds that accumulate in hepatocytes
- Recommend measuring intracellular compound concentration for cell based assays used to provide DILIsym[®] parameter values
 - OCR, ROS production
 - For compounds that are known to have liver:blood ratio \neq 1 (or not known)

HepG2



	extracellular	intracellular
10X accumulation	74.4 µM	744 µM
1x accumulation	74.4 µM	74.4 µM
0.1x accumulation	74.4 µM	7.29 µM