How to obtain biowaivers for clinical trials using PBPK models, two case studies

Maxime Le Merdy Scientist II, Simulations Plus

EUFEMED

<u>Workshop 1:</u> Modeling and simulations, including PBPK to improve the clinical development May 15th 2019



Take home messages:

- FDA is open to proposals of using modeling approaches for bioequivalence (BE), or for new drugs, with the proper justification and model verification
- PBPK models can answer a variety of questions from regulatory agencies
- PBPK is a great tool to understand the interconnection between API properties, formulation attributes and human physiology





- <u>Case Study 1</u>: Crossover trials to show BE after manufacturing changes
- <u>Case Study 2</u>: Long-acting injectables (LAI) generic products development



The Key points from U.S. FDA Workshop:

FLIGHT SIMULATOR: LEARNING HOW TO DEVELOP COMPLEX GENERIC DRUG PRODUCTS

When: Nov 3, 2018 from 9:00 AM to 5:00 PM (ET) Associated with <u>AAPS Community</u>

DOWNLOAD TO YOUR CALENDAR

- FDA is open to proposals using modeling approaches to establish bioequivalence for the "Test" products, as long as these proposals include information about the modeling approach, scientific justification of the proposed approach and in the end model verification.
- Discuss your proposed BE modeling approach through the pre-ANDA development meeting.



Case Study 1:

Crossover trials to show BE after manufacturing changes



Modeling & Simulations Objectives





Proposed Modeling Tasks

- Part 1: Determine the most appropriate absorption/PBPK model for the API across several doses for the non-engineered lots
- Part 2: Assess the effect of particle size on API exposure for the immediate release (IR) formulation
- Part 3: Evaluate predicted bioequivalence of the tablets manufactured with particleengineered (PE) API (narrower particle size distribution) versus the tablets manufactured with non particle-engineered (NPE) API (broader particle size distribution)



Part 1: Building the Baseline Model: Key Modeling Parameters

- BCS Class IV drug
- Neutral compound
- Aqueous solubility = 10 μg/mL
- Significant solubilization by bile salts
- Intermediate lipophilicity
- No food effect

Various Particle Size Used in Clinical Studies

NPE API Lot Number	d10 (µm)	d50 (µm)	d90 (µm)	PE API Lot Number	d10 (µm)	d50 (µm)	d90 (µm)
NPE Lot 1	20	63	173	PE Lot 1	16	40	88
NPE Lot 2	8	179	512	PE Lot 2	20	49	102
NPE Lot 3	15	49	142	PE Lot 3	22	53	108
NPE Lot 4	31	86	348	PE Lot 4	19	39	71
NPE Lot 5	26	78	276	PE Lot 5	17	35	67
NPE Lot 6	9	29	101	PE Lot 6	23	48	93
NPF Lot 7	11	35	114	PE Lot 7	21	44	87
	42	27	424	PE Lot 8	21	45	90
NPE LOT 8	12	37	124	PE Lot 9	24	50	94
NPE Lot 9	10	36	119	PE Lot 10	21	45	89
NPE Lot 10	13	45	138	PE Lot 11	19	42	88
NPE Lot 11	11	35	99	PE Lot 12	22	47	95

API: active pharmaceutical ingredient; d10, d50, and d90: diameter for which 10%, 50%, and 90% (respectively) by volume of the particles are less than this value NPE: non-particle-engineered; PE: particle-engineered

Parameter	Value
CL	0.115 L/h/kg
First pass extraction	17%
Vc	0.324 L/kg
K12	0.26 h ⁻¹
K21	0.1 h ⁻¹

<u>C</u> ompour	nd		Gut	Physiology	-Hum	1	Pharmac	<u>o</u> kinetics	I	Simulation	<u>G</u> raph
Compartmental Parameters								E Ew	orete all un-	absorbed drug at the end of gut trans	ait time.
Propranolol HCI						Va	Values Zero-order gastric emptying				at units
Compartment Data									Enzyme and Transporter	Regional Distributions	
Compartment	Peff	ASF	pН	Transit Time (h)	Volume (mL)	Length (cm)	Radius (cm)	SEF	Bile Salt (mM)		
Stomach	0	0.0	1.30	0.25	48.92	29.19	9.87	1.000	0.0		
Duodenum	0	2.727	6.00	0.26	44.57	14.58	1.56	4.235	2.800		
Jejunum 1	0	2.678	6.20	0.94	166.6	60.26	1.48	3.949	2.330		
Jejunum 2	0	2.675	6.40	0.74	131.0	60.26	1.32	3.489	2.030		
lleum 1	0	2.640	6.60	0.58	102.0	60.26	1.16	3.029	1.410		
lleum 2	0	2.621	6.90	0.42	75.35	60.26	1.00	2.569	1.160		
lleum 3	0	2.589	7.40	0.29	53.57	60.26	0.84	2.109	0.140		
Caecum	0	0.352	6.40	4.36	50.49	13.50	3.45	1.790	0.0		
Asc Colon	0	0.823	6.80	13.07	53.55	28.35	2.45	2.480	0.0		
•									Þ		
C1-C4: 0.06	944	0.43	3028	0.12	147	0.466	32			Qh	(L/min): 1.5
Physiolog	y: Huma	an - Physio	logical - F	asted				•		Percent Fluid in SI: 40	Colon: 10
ASF Mode	el: Optik	ogD Model	ISA/V 6.1					•			

SimulationsP

SCIENICE + SOETWARE = SLICCES

Part 1: Simulation Results for Baseline Models of Non-Engineered Lots



Total simulation t	ime (h): 24	
Result Ol	oserv	Sir	nul
Fa (%)	0	85.90	7
FD _p (%) ———	0	85.90	7
F (%)0	0	71.30	93
Cmax (ng/mL):	391.	2	399.12
Tmax (h):	1.5		2.56
AUC o-inf (ng-h/ml	.) 3563	.7	3739.6
AUC o-t (ng-h/mL):	3139	.1	3702
Cmax Liver (ng/mL)):		531.85



Total simulation time (h): 24						
Result	0bs	erv	Sin	nul		
Fa (%)		0	96.42	2		
FD _p (%)		0	96.42	2		
F (%)0		0	80.03			
Cmax (ng/mL):_	_	2768		3245.8		
Tmax (h):		1.5		2.08		
AUC o-inf (ng-h/	mL)	26290	D	24970		
AUC o-t (ng-h/mI	L):_	22590)	20990		
Cmax Liver (ng/r	nL):			4079.7		



Result	0bserv	Si	nul
Fa (%)	0	85.9	07
FD _D (%)	0	85.9	07
F (%)0	0	71.3	03
Cmax (ng/ml	L):92	6.3	399.12
Tmax (h):	1.9	5	2.56
AUC o-inf (ng-	•h/mL) 75	45.6	8462.2
AUC o-t (ng-h	/mL):_ 63	58.8	7117.3
Cmax Liver (n	g/mL):		1385.9

Same baseline absorption model does a good job of predicting the observed plasma concentrationtime data across the three different doses of the NPE ("old") API lots.



Tistaert, C. AAPS Annual Meeting 2015, Orlando, FL

Part 2: Parameter Sensitivity Analysis (PSA) Around Mean Particle Radius:

<u>Dose Range: 10 – 1000 mg</u>



Part 2: Parameter Sensitivity Analysis (PSA) Around Standard Deviation &

Shape Factor: Dose Range: 10 – 1000 mg



PSA was also run to evaluate changes in particle size standard deviation (assuming mean remained constant) and particle shape factor

Results indicated that there would be insignificant/moderate changes in Fa% across the range of values evaluated



Part 3: Virtual Bioequivalence Trials: Population Simulator

Incorporate measured variability for physicochemical, formulation and PK parameters into Population Simulator

Capture observed variability from existing clinical PK studies







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Part 3: Virtual Bioequivalence Trials: Population Simulator

h/mL]

-Bn]

NUC(0-1)

- Crossover studies simulations for 10 different populations, each with 25 virtual subjects, were run to predict bioequivalence
- 100% <u>passing ratios</u> for C_{max} and AUC were predicted (within the 80-125% limits) between the "new" and "old" API lots (up to 40 µm)



Virtual Bioequivalence Study Simulations

API Lot	PE/NPE	Dose	AUC _∞ (ng.h/mL) (N=250)		(ng.h/mL) C _{max} (ng/mL) N=250) (N=250)	
		(mg)	GM	GMR (90% CI)	GM	GMR (90% CI)
Lot 5	PE	50	4180	113.3	551	139.3
Lot 1	NPE	50	3688	(110.7, 116.1)	395	(136.0, 142.7)
Lot 5	PE	100	8242	103.0	551	106.4
Lot 3	NPE	100	8001	(100.9, 105.1)	395	(104.3, 108.6)
Lot 5	PE	300	24998	102.2	3118	100.0
Lot 2	NPE	300	24460	(99.8, 104.6)	3117	(97.7, 102.4)
Lot 5	PE	100	8242	98.2	1068	95.1
Lot 4	NPE	100	8395	(96.2, 100.2)	1123	(93.2, 97.0)
Lot 5	PE	300	24998	101.9	3118	98.3
Lot 4	NPE	300	24525	(99.8, 104.1)	3171	(96.3, 100.4)

API: active pharmaceutical ingredient; AUC_{∞} : area under the plasma concentration-time curve from time 0 to infinite time; CI: confidence interval; C_{max} : maximum observed plasma concentration; GM: geometric mean; GMR: geometric mean ration; NPE: non-particle-engineered; PE: particle-engineered



Tistaert, C. AAPS Annual Meeting 2015, Orlando, FL

<u>Summary</u>

- A mechanistic, physiologically-based absorption/PK model was constructed in GastroPlus and validated across three dose levels (50, 100, and 300 mg) using *in vivo* data collected from tablets manufactured with non-particle engineered API.
- Parameter sensitivity analysis showed that mean particle size would be the main property that determines whether formulations are likely to be bioequivalent, regardless of dose.
- Virtual bioequivalence trial simulations showed, that for a sufficiently powered study the population-derived C_{max} and AUC values would be bioequivalent between the tablets manufactured with non-particle engineered (NPE) vs. new particle engineered (PE) API, up to 40 μm particle size, regardless of the dose.
- Regulatory agencies approved the sponsor's biowaiver application



Case Study 2:

Long-acting injectables generic product development



Goal of the project

 Long-acting injectable (LAI) formulations include biodegradable injectable microspheres and in-situ gelling implants. Compendial in *vitro* release methods for these complex formulations are not well developed, and demonstration of BE for these products can be challenging.

	Contains Nonbinding Recommendations Draft Guidance on Naltrexone	
Active Ingredient:	Naltrexone	
Dosage Form; Route:	Extended-release suspension; intramuscular	
Recommended Studies: 1. Type of study: In vi Design: Design:	One study vo single-dose fasting	The FDA consider these
Strength: 380 mg/vi Subjects: Healthy m Additional comment test/reference (T/R) fall within the limits	al (dose: 380 mg) hales and nonpregnant females, general population the 90% confidence intervals of the geometric mean ratios for the metrics (C _{max} , AUC ₁₋₁₀ , AUC ₁₀₋₂₈ , and AUC _{0-∞}) should of 80-125%	"complex products"

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Aims of the project

Introduction to Complex Products and FDA Considerations

Xiaohui (Jeff) Jiang, PhD Deputy Director Division of Therapeutic Performance Office of Research and Standards Office of Generic Drugs Center for Drug Evaluation and Research, FDA

In 2015, Simulations Plus and a major pharmaceutical company received a grant from the FDA to improve the pre-existing model for in *vitro in vivo* correlation (IVIVC) within GastroPlus[®].

Bridging in vitro and FDA in vivo studies In vivo performance GastroPlus™ For LAI Drug Discovery In vitro testing and Development 26 SimulationsPlus **S**+

https://www.fda.gov/media/108937/download

In vitro-In vivo correlation: Mechanistic absorption model

• Working definition:

"A predictive mathematical treatment describing the relationship between an *in vitro* property of a dosage form (e.g., the rate or extent of drug release) and a relevant *in vivo* response (e.g., plasma concentration-time data)"

FDA Guidance for Industry Extended Release Solid Oral Dosage Forms: Development, Evaluation, and Application of *In Vitro/In Vivo* Correlations (1997)

<u>Method:</u>



- Inputs:
 - Physiological parameters
 - Drug properties (solubility, Peff, logP, pKa, etc.)
 - PK data
 - In vitro dissolution profile



A model that combines all available *in silico*, *in vitro* and *in vivo* information and provides:

- *in vivo* dissolution, absorption and bioavailability vs. time profiles
- Description of site dependent absorption
- Description of tissue contributions to first pass extraction



In vitro-In vivo correlation (IVIVC): Mechanistic absorption model



Find the correlation between the deconvoluted in vivo release and in vitro dissolution profiles



Predict the plasma concentration-time profile using the IVIVC and in vitro dissolution curve



Complex *in vivo* **Profile**

The *in vivo* release profiles for LAIs are often complex and cannot be described well with a single or double-Weibull function (typically sufficient for deconvolution of *in vivo* release profiles for standard oral formulations). **This issue was addressed by adding triple-Weibull function**.

Simulated Cp-time profiles after SC injection of one LAI formulation in rat. The *in vivo* release profile was fitted as double-Weibull (top) and triple-Weibull (bottom) function with 3 objective function weighting schemes



🖉 Weibull Controlled Release Profile	- 🗆 ×
File	
Comments: ['In Vitro Amt [mg] =	\$
$\label{eq:DoseReleased} \begin{split} & \% DoseReleased = Max \times \left(1 - f_1 exp\left[\frac{-(t-T)^{b_1}}{A_1}\right] - f_2 exp\left[\frac{-(t-T)^{b_2}}{A_2}\right] - f_3 exp\left[\frac{-(t-T)^{b_3}}{A_3}\right]\right) \end{split}$	
Weibull Parameters Fit I (time lag) (hrs): I Select Weibull Function:	[%] p
Ma <u>x</u> (total released) (%): 100 Double Weibull	- 0 - 0 - 0 - 0 - 0 - 0 - 0 - 0 - 0 - 0
Phase 1Phase 2Fitf (fraction):1 \Box \Box \Box \Box A (time scale) (hrs^b):1 \Box 1 \Box	B.
	0 Time [hrs]
Find Initial Estimates Fit Weibull Function Cancel OK	



Flexibility of the Weibull Function?

GastroPlus offers single-, double-, and triple-Weibull function for optimization of *in vivo* release profile, which cover a wide variety of release profile shapes.



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Optimization Target Criteria

When fitting the *in vivo* release profile against the entire observed Cp-time profile, the error on Cmax is often higher than allowed by the IVIVC criteria due number of other concentration points outweighing the contribution of the single Cmax value.

This issue will be addressed by adding option to include additional weight on Cmax during deconvolution in IVIVCPlus[™] module.

Simulated Cp-time profiles for 3 naltrexone LAI microsphere formulations.

Top row shows deconvolution results with target observed Cp-time profile,

Bottom row shows deconvolution results with additional weight added to Cmax value.



Insufficient *in vitro* Sampling

Insufficient *in vitro* sampling may cause difficulties matching the shape of the predicted Cp-time profile even with an otherwise valid IVIVC (correct prediction of Cmax and AUC).

This issue will be addressed by adding an option to use interpolated *in vitro* profile in IVIVCPlus module.

IVIVC for huperzine A LAI microspheres. Plots on the left showing IVIVC created are from 2 LAI formulations using only the measured in vitro points (top) and interpolated in vitro points at the early timepoints (bottom). Plots on right the are showing corresponding predicted and observed Cp-time profiles for third LAI formulation.



Variability in IVIVC Across Formulations

The *in vivo* release profiles were successfully deconvoluted using a triple-Weibull function for all formulations; however, the differences in the deconvoluted *in vivo* release profiles are not accurately captured by the differences in the measured *in vitro* release profiles.

Literature search was performed to identify possible mechanisms responsible for differences between *in vitro* and *in vivo* release for LAI formulations.

Top: Simulated Cp-time profiles for 4 orntide LAI microsphere formulations after successful deconvolution of *in vivo* release profiles.

Bottom: Deconvoluted *in vivo* release profiles (left) and IVIVC plot (right) for 4 orntide LAI microsphere formulations.



The Tissue Response to PLGA Microsphere Administration

- The tissue response to PLGA microsphere administration can be divided into three phases:
- I. Acute phase of the inflammatory response
- II. Onset of the chronic phase of inflammation
- III. Fibroblasts infiltration and collagen deposition





The temporal variation in the three phases of inflammatory response resulting from administration of biodegradable microspheres



Anderson et. al., Advanced Drug Delivery Reviews 64 (2012), 2012

Quantitative Evaluation and Modeling of the

Depot Infiltration: Immune Cell Layer



Darville et al, Toxicologic Pathology, 2016, Vol. 44(2) 189-210



Intramuscular: Immune Cell Layer



Outcomes

□ Implementing *in vitro* drug release from PLGA microspheres in DDDPlus.

Additional functionalities added to the IVIVC module in GastroPlus

- **Triple Weibull** function for deconvolution
- □ The capability of perfuming IVIVC using interpolated data
- □ Adding **Shifting and Scaling** function to the list of correlation functions
- □ Adding users the capability of choosing different optimization function as part of deconvolution
- Adding the option for **Setting the intercept to zero** to Deconvolute then Correlate.
- Adding Intramuscular and Subcutaneous route of administration to the IVIVC module in GastroPlus.

□ Immune Cell Layer (ICL) Model



Summary

- The mechanistic absorption model-based IVIVC was improved to be able to successfully deconvolute the *in vivo* dissolution profile for LAI based on clinical PK data.
- Correlation process was improved to better linked in *vitro* and *in vivo* release profiles
- Other mechanism seem to be important *in vivo*: inflammation and immune cell infiltration
- Regulatory agencies have access to these models and will use them for BE guidance development and reviews



Publications/Presentations

Jul 16, 2017-CRS Simulation of in vitro Dissolution and Degradation of Orntide-loaded PLGA Microspheres Mullin J, van Osdol W, Lukacova V, Woltosz WS, Bolger MB

□ Nov 2016-AAPS

Development of an In Vitro Mechanistic Model that Describes Drug Release from Risperidone Long Acting Injectable Microspheres

James Mullin; Viera Lukacova; Walter Woltosz; Michael B. Bolger

□ Nov 14, 2017-AAPS

Development of In Vitro-In Vivo Correlation for Long Acting Injectable Microsphere Formulations Shahraz A, Mullin J, Spires J, Lukacova V, Bolger MB, Woltosz WS

□ Poster accepted for ACoP10

Modeling and Simulation of the Local Tissue Response to Long-acting Injectable Formulations

Azar Shahraz, James Mullin, Viera Lukacova



GastroPlus® PBPK Consulting: Regulatory Submissions

- Since 2016, our consulting team has built PBPK models for over 110 applications, some of which were used to support submissions to various regulatory agencies:
 - Preclinical development and First-in-Human predictions
 - Formulation optimization
 - DDI predictions
 - Virtual bioequivalence trial simulations
 - Pediatric population simulations and dose projections
 - Food effect modeling
 - Parent-metabolite and prodrug PBPK modeling
 - Pulmonary/dermal/oral cavity product assessment
 - Mechanistic IVIVCs to define product specifications



Key Points to be Addressed to Regulators

- Objective and intended regulatory purpose of the PBPK modeling
- Sufficient background information to place the PBPK modeling in its context in the clinical development of the drug
- Model validation and explicit/systematic discussion of the assumptions made in the submitted drug model and analysis, i.e. supportive data and biological plausibility and impact of the assumptions on the model and the outcome
- Sensitivity analysis, especially on the parameters that were fitted in the model



Questions?

