

# Quantitative Modeling Uncovers a Potential Limitation in the Putative Mechanism of CCl<sub>4</sub> Hepatotoxicity

Lisl K. M. Shoda; Scott Q. Siler; Paul B. Watkins; Brett A. Howell

The Hamner - UNC Institute for Drug Safety Sciences, The Hamner Institutes for Health Sciences, Research Triangle Park, NC

## Abstract

Drug-induced liver injury (DILI) is one of the leading causes of drug development failures and drug withdrawals. DILIsym<sup>®</sup> is being developed to identify and mitigate DILI risk through in silico analysis of compounds. The DILIsym<sup>®</sup> representation of the innate immune response was initially based on acetaminophen (APAP) data. Carbon tetrachloride (CCl<sub>4</sub>), a compound with hepatotoxic similarities to APAP, was simulated for further evaluation of the innate immune response to liver injury. DILIsym<sup>®</sup> simulation results for CCl<sub>4</sub> pharmacokinetics were consistent with published PK data. CCl<sub>4</sub> is generally thought to induce hepatotoxicity via a free radical metabolite which drives lipid peroxidation. However, when CCl<sub>4</sub> was simulated via DILIsym<sup>®</sup>, free radical generation and lipid peroxidation to levels consistent with the public literature were insufficient to drive hepatotoxicity. Analysis demonstrated that saturation of the metabolic pathway generating the free radical limited the extent of lipid peroxidation and thus the extent of cell death. In comparison, the APAP metabolic pathway has a greater dynamic range, resulting in higher levels of lipid peroxidation and cell death. Papers were identified in which lipid peroxidation independent mechanisms of cell death were suggested, including lipid peroxidation independent mitochondrial toxicity. The CCl<sub>4</sub> representation was modified to induce mitochondrial electron transport chain (ETC) inhibition. Concurrent induction of lipid peroxidation and ETC inhibition mechanisms permitted reconciliation of simulated hepatotoxicity with published data. For example, >3x ALT elevations were simulated at doses <50 mg/kg, with the highest ALT elevations (>1000 U/L) at doses >100 mg/kg in mice. Simulated CCl<sub>4</sub> was then used to evaluate and further refine the innate immune response. In summary, quantitative analysis of CCl<sub>4</sub> in DILIsym<sup>®</sup> not only permitted evaluation of the innate immune response, but also suggested a limitation in the putative primary mechanism of hepatotoxicity.

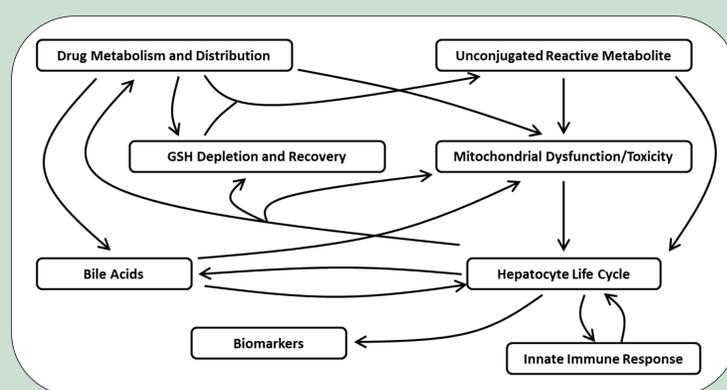
## Introduction

DILIsym<sup>®</sup> is a multi-scale mechanistic model of drug-induced liver injury (DILI). The model contains a physiologically-based pharmacokinetic model of drug distribution and metabolism in the liver and several mechanisms of toxicity. Acetaminophen (APAP) has been used as an exemplar compound for reactive metabolite mediated oxidative stress and hepatotoxicity based on the wealth of available data.

The immune response has been implicated in various forms of DILI. As a first step to incorporating immune responses in DILI, macrophage participation in APAP overdose has been added to DILIsym<sup>®</sup>. To increase confidence that the macrophage model was adaptable to other compounds, carbon tetrachloride (CCl<sub>4</sub>) was tested. Similar to APAP, the liver metabolizes CCl<sub>4</sub> to a reactive metabolite (RM). RM-generated oxidative stress is the putative primary mechanism of hepatotoxicity. There are also published data illustrating the recruitment of the immune response in CCl<sub>4</sub> hepatotoxicity.

We re-parameterized the DILIsym<sup>®</sup> compound physiologically based pharmacokinetic (PBPK) model and compound metabolism model to represent CCl<sub>4</sub>. We selected RM-induced oxidative stress as the mechanism of hepatotoxicity and then ran simulations to explore the relationship between CCl<sub>4</sub> hepatotoxicity and recruitment of the immune response.

## Overview of DILIsym<sup>®</sup>



## Dual Mechanism CCl<sub>4</sub> Simulation Results

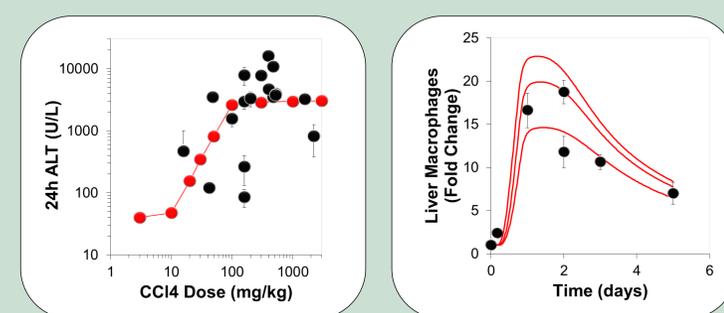


Fig 3a. Adding ETC inhibition permits CCl<sub>4</sub> simulations, red, consistent with the observed hepatotoxicity, black [8,10-21]

Fig 3b. With CCl<sub>4</sub> hepatotoxicity, simulations of macrophage accumulation are consistent with published data [8, 9]

## Initial CCl<sub>4</sub> Simulation Results

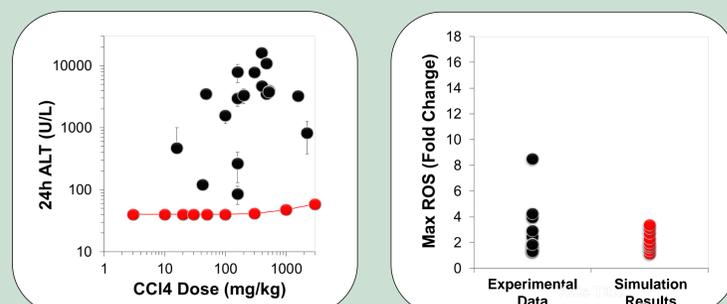


Fig 1a. Initial representation of CCl<sub>4</sub>, red, fails to recapitulate observed mouse hepatotoxicity, black [8, 10-21]

Fig 1b. Simulated CCl<sub>4</sub> (3-3000 mg/kg) generates oxidative stress but less than simulated APAP (Fig 2a) [15, 22-32]

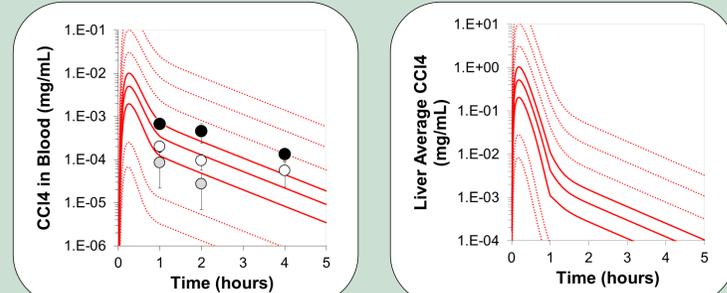


Fig 1c. PBPK simulations (1-3000 mg/kg). Solid red lines are the result of dosing as reported [1] and are consistent with data.

Fig 1d. Liver exposure increases with higher doses of CCl<sub>4</sub> (simulations as in Fig 1c)

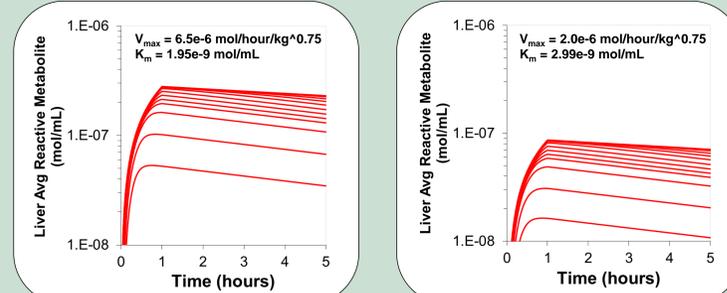
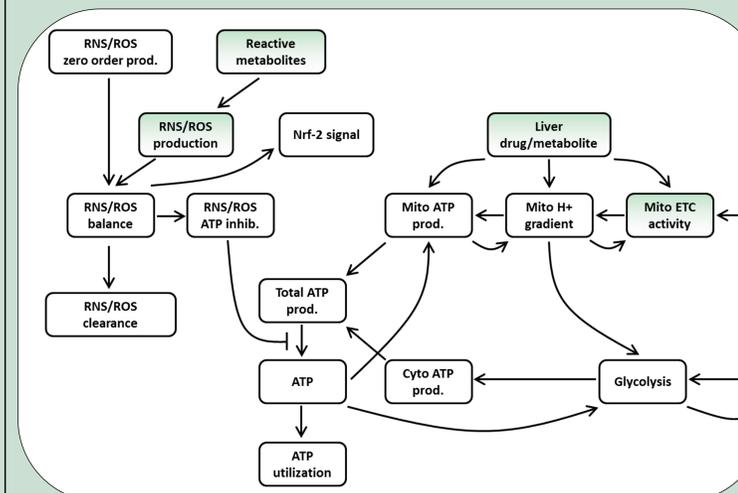


Fig 1e. RM generation becomes saturated at higher doses of CCl<sub>4</sub> (1-3000 mg/kg)

Fig 1f. Saturation persists with alternate published values for CCl<sub>4</sub> metabolism

## Mitochondrial Dysfunction/Toxicity

To explore the possible contribution of mitochondrial dysfunction in CCl<sub>4</sub> hepatotoxicity, a second mechanism, CCl<sub>4</sub> RM-induced inhibition of the electron transport chain (ETC) activity was introduced.



## Contrast to Acetaminophen

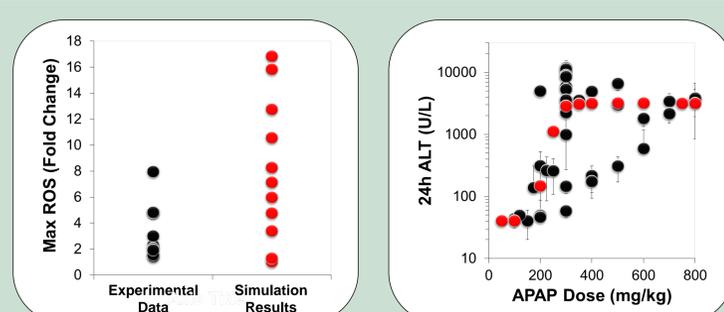


Fig 2a. In contrast to CCl<sub>4</sub>, APAP RM generation is not limiting and can lead to extensive oxidative stress [33-42]

Fig 2b. In contrast to CCl<sub>4</sub>, APAP RM-mediated oxidative stress is sufficient to simulate, in red, dose-dependent hepatotoxicity consistent with published data, in black [35, 43-58]

## Methods

- We simulated mice given single doses of CCl<sub>4</sub> orally or intraperitoneally in aqueous or oil-based vehicle
- CCl<sub>4</sub> tissue distribution, fraction unbound, and clearance were estimated from the literature [e.g., 1-3] and tuned to match published time course profiles [1] (Fig 1c)
- CCl<sub>4</sub> metabolism was simplified to represent generation of the reactive metabolite (RM), trichloromethyl peroxy radical, implicated in CCl<sub>4</sub> hepatotoxicity [4]
  - RM generation was governed by V<sub>max</sub> and K<sub>m</sub> values, taken from the published literature [1]
  - Suicide inhibition of the CYP450 pathway was simplified to represent 95% inhibition after 1 hour exposure in mice [1]
- The same approach was taken for the exploration of rat CCl<sub>4</sub> responses. *Rat simulations not shown.*

## Results and Analysis

- In initial simulations, CCl<sub>4</sub>-generated oxidative stress was unable to reproduce dose-dependent hepatotoxicity (Fig 1a, b)
- Analysis demonstrated that hepatotoxicity was limited by saturation of CCl<sub>4</sub> metabolism (Fig 1e)
  - Use of alternate V<sub>max</sub> and K<sub>m</sub> values for CCl<sub>4</sub> metabolism [5] led to saturation of RM generation at a lower level and did not change the hepatotoxicity outcomes (Fig 1f)
  - Further literature review suggested the possibility of direct mitochondrial effects [6, 7]
- CCl<sub>4</sub>-mediated mitochondrial dysfunction, i.e., electron transport chain (ETC) inhibition, permitted the simulation of CCl<sub>4</sub> hepatotoxicity (Fig 3a)
- Simulated macrophage accumulation was consistent with experimental observations [8, 9] (Fig 3b)

## Conclusions

- Quantitative modeling and simulation of CCl<sub>4</sub> suggests saturation of CCl<sub>4</sub> metabolism could limit RM-mediated oxidative stress and resultant hepatotoxicity
- Data on mitochondrial liability may indicate an underappreciated mechanism of toxicity
- Once CCl<sub>4</sub> dose-dependent hepatotoxicity was simulated, a reasonable accumulation of macrophages could be confirmed.

## Acknowledgements

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## References

- Fisher, J. et al. PBPK modeling of the metabolic interactions of carbon tetrachloride and tetrachloroethylene in B6C3F1 mice. *Environ. Toxicol. Pharmacol.* 16, 93-105 (2004).
- Endo, S. & Goss, K.-U. Serum albumin binding of structurally diverse neutral organic compounds: data and models. *Chem. Res. Toxicol.* 24, 2293-2301 (2011).
- Sanzgiri, U. Y., Srivatsan, V., Muralidhara, S., Dallas, C. E. & Bruckner, J. V. Uptake, distribution, and elimination of carbon tetrachloride in rat tissues following inhalation and ingestion exposures. *Toxicol. Appl. Pharmacol.* 143, 120-129 (1997).
- U.S. Environmental Protection Agency. TOXICOLOGICAL REVIEW OF CARBON TETRACHLORIDE (CAS No. 56-23-5). (2010). at <http://www.epa.gov/iris/toxreviews/0020tr.pdf>
- Thrall, K. D. et al. Comparative metabolism of carbon tetrachloride in rats, mice, and hamsters using gas uptake and PBPK modeling. *J. Toxicol. Environ. Health A* 60, 531-548 (2000).
- Albano, E. et al. Mechanisms responsible for carbon tetrachloride-induced perturbation of mitochondrial calcium homeostasis. *FEBS Lett.* 192, 184-188 (1985).
- Knockaert, L. et al. Carbon tetrachloride-mediated lipid peroxidation induces early mitochondrial alterations in mouse liver. *Lab. Invest. J. Tech. Methods Pathol.* 92, 396-410 (2012).
- Minamino, T. et al. Thromboxane A<sub>2</sub> receptor signaling promotes liver tissue repair after toxic injury through the enhancement of macrophage recruitment. *Toxicol. Appl. Pharmacol.* 259, 104-114 (2012).
- Karimk, K. R. et al. Hepatic recruitment of the inflammatory Gr1+ monocyte subset upon liver injury promotes hepatic fibrosis. *Hepatology*. Baltimore, Md 50, 261-274 (2009).

References 8-58 available on request.