# Mechanistic Modeling of Kidney-Injury Molecule 1 (KIM-1) as a biomarker for Cisplatin-Induced Acute Kidney Injury

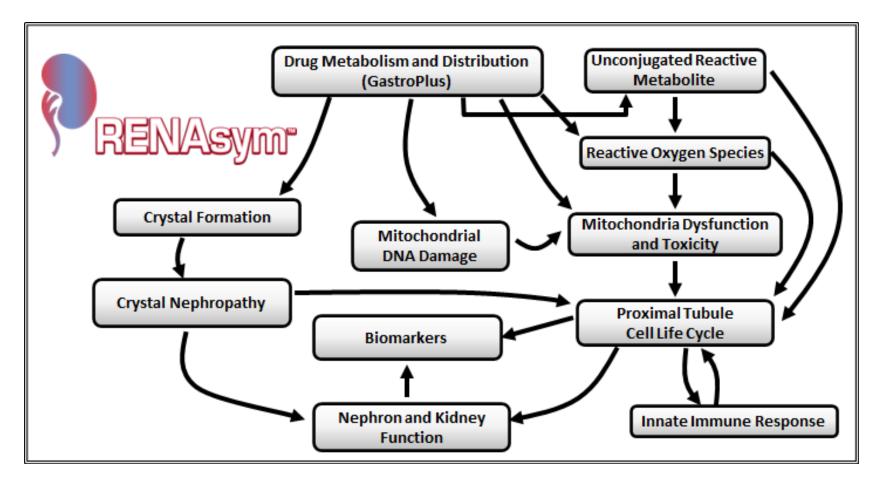
### INTRODUCTION

- Kidney Injury Molecule 1 (KIM-1) is a specific and sensitive biomarker for drug-induced acute kidney injury (AKI) prediction
- Despite growing interest in clinical use of KIM-1 as a key biomarker for AKI diagnosis, a mechanistic model of KIM-1 that accurately predicts the kinetics of KIM-1 is still lacking.
- Unlike normal conditions where urinary Kim-1 is not detectable, it is significantly expressed during acute kidney injury. Kim-1 was detected at high levels in proliferating bromodeoxyuridinepositive and dedifferentiated vimentin-positive epithelial cells in regenerating proximal tubules (Ichimura 1998).
- We developed a mechanistic model of KIM-1 as part of a quantitative systems toxicology (QST) model to predict urinary KIM-1 in rats, mice and human treated with cisplatin.
- Our objective is to characterize cisplatin-induced injury of the renal proximal tubular epithelial cells (RPTEC) and biomarker responses using Kim-1 in vivo studies

### **METHODS**

- A quantitative systems toxicology model, RENAsym<sup>™</sup>, recently developed for drug induced acute kidney injury by DILIsym Services. This model embodies key cellular injury mechanisms and renal hemodynamics.
- A mechanistic model of KIM-1 was developed within the framework of RENAsym<sup>™</sup>.
- The KIM-1 model represents the early shedding of KIM-1 arising from the loss of brush borders during sub-lethal injury of RPTEC followed by the marked expression of KIM-1 in dedifferentiated cells in regenerating proximal tubules.
- The model is integrated with the RENAsym to capture RPTEC injury and regeneration following toxic renal injury.

# **RENAsym QST Model**



- comes from existing Kim-1

- decrement
- cellular death

- preclinical/clinical data

#### **Rats**

#### **Humans**

- 2015)

#### 

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## RESULTS

• Kim-1 is considered an inducible biomarker that significantly upregulated during AKI and released into the urine • In our model, the production of Kim-1 mainly originates from upregulated Kim-1 in dedifferentiated cells and early shedding of Kim-1

1 + Existing Kim + Kim + Wim + Kim + Kim

1. The basal Kim-1 production at steady state is set to be very low in line with the basal shedding rate of Kim-1 reported in control healthy individuals (Peters 2011)

2. Urinary Kim-1 is reported before dedifferentiation process begins, and it may be attributed to proximal tubule brush border loss - An existing level of Kim-1 is assumed to be sitting on PTCs and its shedding rate into the urine is determined based on ATP

3. Kim-1 is markedly upregulated in regenerated PTCs and mechanistically linked with the flux of dedifferentiated cells in RENAsym - Increase in dedifferentiated cells is associated with increase in hepatocyte growth factor (HGF) following apoptotic or necrotic

# Model Calibration

• Calibration of model compares well to available preclinical and clinical data

– A PBPK model built in GastrPlus9.8 to determine the kidney intracellular concentration of cisplatin and was further verified with clinical and preclinical data

- Cisplatin exposure from PBPK model was combined with drug toxicity parameters obtained by parameterization of in vitro assay results in Renal Proximal Epithelial Tubular Cells (RPTEC) and translated from *in vitro* to *in vivo* for each species

 Cumulative urinary Kim-1 was obtained from simulations and urinary Kim-1 fold change for every day was calculated, and calibrated Kim-1 timing/magnitude for cisplatin-mediated injury with

– Kim-1 model including upregulation rate constant for PTC dedifferentiated cells and early shedding rate constant were adjusted to recapitulate human, rat and mouse Kim-1 data

Rats treated with cisplatin at 2.5 mg/kg were simulated in RENAsym

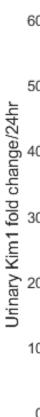
- Simulation results show that the shedding of existing Kim-1 results in 20 fold-increase of urinary Kim-1 around day 4 (orange line)

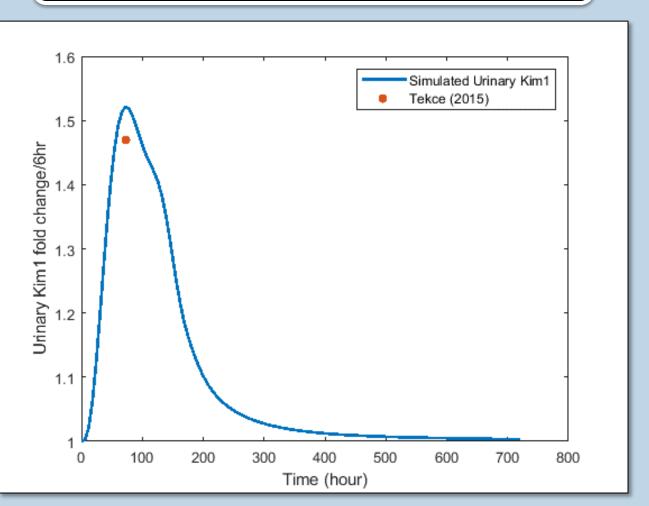
– Urinary Kim-1 fold change peaks between day 5 and day 6 mainly due to Kim-1 upregulation - The kinetics of urinary Kim-1 fold change from simulations agrees well with the observed Kim-1 profile (Gebremichael 2018 and Vaidya 2006)

– Humans treated with 4hr IV infusion of cisplatin at 75 mg/m2 were simulated in RENAsym – Simulated urinary Kim1 fold change per 6hr well captures the observed urinary Kim-1 peak (Tekce

Mice treated with cisplatin at 20 mg/kg were simulated in RENAsym

- Simulated urinary Kim1 fold change per 24hr shows a good agreement with the peak in the observed data for mice treated at the same dose (Oh 2017)





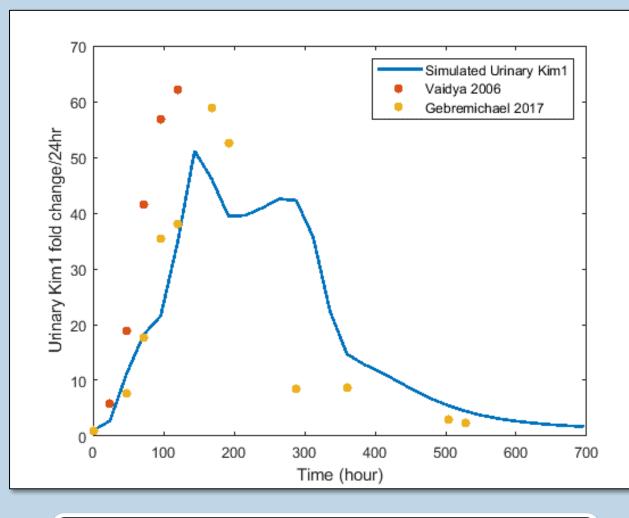
**Kim-1 Mechanistic Model Cellular Toxicity** (ROS, Mito Dysf., Crystal Neph) Early Injury (Brush Border Loss) Cell Death (apoptosis/necrosis Dedifferentiation **Kim-1 Upregulation** Urinary Kim-1

# — Simulated Urinary Kim1 Simulated Urinary Kim1 w/o upregulation 400 200 500 300 600 Time (hour)

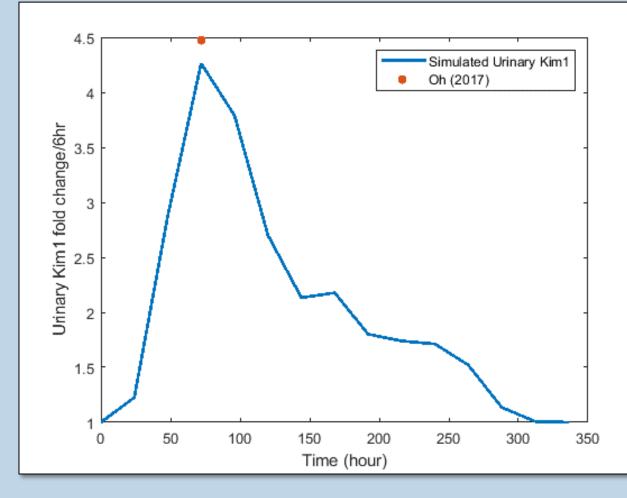
Rat cisplatin at 2.5 mg/kg

# Human cisplatin at 75 mg/m2

Rat cisplatin at 2.5 mg/kg



Mouse cisplatin at 20 mg/kg



### CONCLUSION

#### Developed a mechanistic model of urinary Kim-1 biomarker response to characterize cisplatin-mediated acute kidney injury

- Kim1 model recapitulate the Kim-1 profile in rats treated with cisplatin as well as the Kim-1 peak in humans and mice
- Kim-1 upregulation is determined to be the main contributor to urinary Kim-1 during AKI
- Using dedifferentiation as the driving signal for Kim-1 upregulation, the timing of predicted Kim-1 peak aligns with the clinical and preclinical data
- As dedifferentiation starts with a delay, we could capture the early increase in urinary Kim1 by assuming the shedding of existing Kim-1 sitting on PTCs

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