ABSTRACT

OBJECTIVE: To predict species differences in CKA-mediated hepatotoxicity using DILIsym[®], a mechanistic model of druginduced liver injury (DILI)

METHODS: Inhibitory effects of CKA on bile acid (BA) and bilirubin transporters were assessed using transporteroverexpressing vesicles and cells. CKA-mediated oxidative stress and mitochondrial dysfunction were determined in cells. These in vitro data were used to define HepG2 hepatotoxicity parameters and combined with PBPK sub-model simulations of hepatic compound exposure to predict DILI in DILIsym. Previously constructed human and rat simulated populations (SimPops[™]) that incorporate variability in the aforementioned toxicity mechanisms were utilized to determine the impact of inter-individual variability upon administration of single CKA doses of 900mg and 500mg/kg, respectively.

RESULTS: CKA induces oxidative stress (oxidative stress production rate constant=7278mL/mol/hr human, 9705 mL/mol/hr rat) and inhibits mitochondrial electron transport chain (ETC) flux (ETC inhibition coefficient=14.2 mM human, 1.42 mM rat). CKA inhibits human BSEP (IC_{50} =129.7 µM), rat Bsep ($IC_{50}=94 \mu M$), human MRP3 ($IC_{50}=11.2 \mu M$), human NTCP (IC_{50} =19.5 µM), rat Mrp2 (IC_{50} =68.5 µM), and human OATP $(IC_{50}=0.84 \ \mu M)[1]$. CKA was modeled as a noncompetitive inhibitor of BSEP/Bsep and MRP3/Mrp3 and a competitive inhibitor of NTCP/Ntcp. Due to lack of data, IC₅₀ values for NTCP/Ntcp, MRP3/Mrp3, MRP2/Mrp2, and OATP/Oatp were assumed the same across species. Human SimPops predicted modest increases <1.5x upper limit of normal (ULN) in serum ALT, recapitulating the clinical data. Rat SimPops predicted ALT elevations >3xULN in 36.4% of the population, slightly underpredicting the 75% observed in preclinical trials. DILIsym recapitulated preclinical observations in bilirubin increase due to both DILI and bilirubin transporter inhibition. Rat SimPops predicted increases in total bilirubin >2xULN for 100% of the population, mirroring preclinical data. Conversely, no significant increases in bilirubin were observed in human SimPops, consistent with clinical observations.

CONCLUSION: Using in vitro data to determine toxicity parameters, DILIsym accurately predicted CKA hepatotoxicity in rats and not in humans, consistent with observed preclinical and clinical data.

INTRODUCTION

- CKA is an antirheumatic drug intended as a treatment for rheumatoid arthritis and chronic obstructive pulmonary disorder.
- CKA produced dose-dependent hepatotoxicity signals in preclinical species (rats), where an increased -- but reversible -- level of liver enzymes was observed in single, supratherapeutic dose studies. Hepatotoxicity was not observed in human trials.
- In vitro experiments determined CKA to inhibit bile acid transporters, induce production of reactive oxygen species, and inhibit the mitochondrial electron transport chain activity.
- DILIsym is a systems pharmacology model of DILI that incorporates drug/metabolite disposition, bile acid physiology and pathophysiology, the hepatocyte life cycle, and liver injury biomarkers.





MECHANISTIC MODELING WITH DILISYM® PREDICTS SPECIES DIFFERENCES IN CKA VIA MULTIPLE HEPATOTOXICITY MECHANISMS Christina Battista^{1,2}, Kyunghee Yang², Jerome Mettetal³, Simone Stahl⁴, Paul Watkins^{1,2}, Brett Howell², Scott Siler² ¹Division of Pharmacotherapy and Experimental Therapeutics, Eshelman School of Pharmacy, The University of North Carolina at Chapel Hill, Chapel Hill, NC; ²DILIsym Services, Inc., Research Triangle Park, NC; ³AstraZeneca, Cambridge, MA; ⁴AstraZeneca, Cambridge, UK



constant and ROS production constant parameters were optimized to data using MITOsym and DILIsym, respectively.



CONCLUSION

• DILIsym modeling based on *in vitro* data for the aforementioned toxicity mechanisms qualitatively predicted species differences in CKA hepatotoxicity.

• ETC inhibition is statistically the most potent toxicity mechanism with bile acid transporter inhibition also contributing to ALT elevations. ROS generation has a minimal effect on CKA-mediated hepatotoxicity.

Quantitative systems pharmacology models (QSP) can be used to evaluate DILI mechanisms and investigate observed species differences.



THE UNIVERSITY of NORTH CAROLINA at CHAPEL HILL

	Simulations			Preclinical / Clinical Trials		
Species	rat	rat	human	rat	rat	hum
Dose	200 mg/kg	500 mg/kg	900 mg	200 mg/kg	500 mg/kg	900 r
Population size	n=294	n=294	n=285	n=8	n=4	n=8
ALT > 3x ULN (%)	2.4	36.4	0	25	75	16.
ALT > 5x ULN (%)	0	20.1	0	0	50	0
ALT > 10x ULN (%)	0	7.8	0	0	25	0
TBIL > 1.5x ULN (%)	100	100	0	100	100	0
TBIL > 2x ULN (%)	0	83	0	100	100	0
Hy's Law cases (%)	-	-	0	-	-	0



Each mechanism was modeled as the sole mechanism (left figure, top row) to study its individual effect on toxicity. ETC inhibition appears to be the main contributor to CKA-mediated rate hepatotoxicity. Two mechanisms are modeled at a time (left figure, bottom row). ETC inhibition and BA transporter inhibition together produce a greater peak plasma ALT, suggesting BA transporter inhibition as the secondary contributor to hepatotoxicity. The multiple regression analysis found the basal value of ETC flux, a parameter in the mitochondrial toxicity submodel, to be most closely correlated with simulated ALT elevations (right).

ACKNOWLEDGEMENTS

- This project was supported in part by an appointment to the Research Participation Program at the Center for Drug Evaluation and Research, U.S. Food and Drug Administration, administered by the Oak Ridge Institute for Science and Education through an interagency agreement between the U.S. Department of Energy and FDA.
- The current research was supported by members of the DILI-sim Initiative.

from (TBIL) fold

> qualitatively species ALT



METHODS

Determining CKA toxicity mechanism parameters from in vitro experiments MITOsym[®], a mitochondrial modeling platform used to mimic in vitro experimental results², was used to determine parameter values for CKA-mediated mitochondrial ETC inhibition by fitting simulated results to Seahorse XF96 basal respiration data for HepG2 cells exposed to CKA. Conversion factors translated predicted results from HepG2 cells to human and rat hepatocytes. The ROS production constant parameter was found by sweeping through potential values until the DILIsym "in vitro-like" simulation yielded ROS effects in line with the data. The BSEP/Bsep IC₅₀ for CKA was used to represent the inhibition constant (Ki) in DILIsym (Ulloa et al., provided by Astra Zeneca, unpublished). Further studies performed in transporter-overexpressing vesicles determined IC₅₀ values for MRP3, MRP4, and NTCP which were used for both species.

Physiologically-based pharmacokinetic (PBPK) model development Human and rat PBPK models were developed in DILIsym using available in vitro and in vivo pharmacokinetic data.

Simulation of CKA responses Previously constructed SimPops that include variability in species-specific parameters and the BA, ROS, and mitochondrial submodels within DILIsym were used to predict the hepatotoxic effects of CKA at the population level. For humans, single doses of 600 and 900 mg were simulated for a period of 96 hours following the administration of CKA. Rat simulations were run for 72 hours after single doses of 200 or 500 mg/kg.

Multiple Regression Analysis To identify the most important parameters associated with DILI in rats, a multiple regression analysis was performed with maximum ALT as the dependent variable. The 34 parameters varied in the rat SimPops were normalized and used as the independent variables. Statistical analyses were performed using JMP 11 (SAS, Cary, NC).



DILIsym[®] Overview

A general overview of DILIsym and its submodels. Topics outlined in red indicate active submodels used in modeling CKA-mediated effects in human and rat populations. Mechanistic in vitro toxicity data were used in the intracellular bile acids, reactive oxygen species, and mitochondria dysfunction and toxicity submodels to predicitDILI outcomes.

REFERENCES

Ulloa, J.L. et al. NMR Biomed 26(10): 1258—1270. 2013. 2. Yang, Y. et al. *Pharm Res* 32(6): 1975—1992. 2015.

