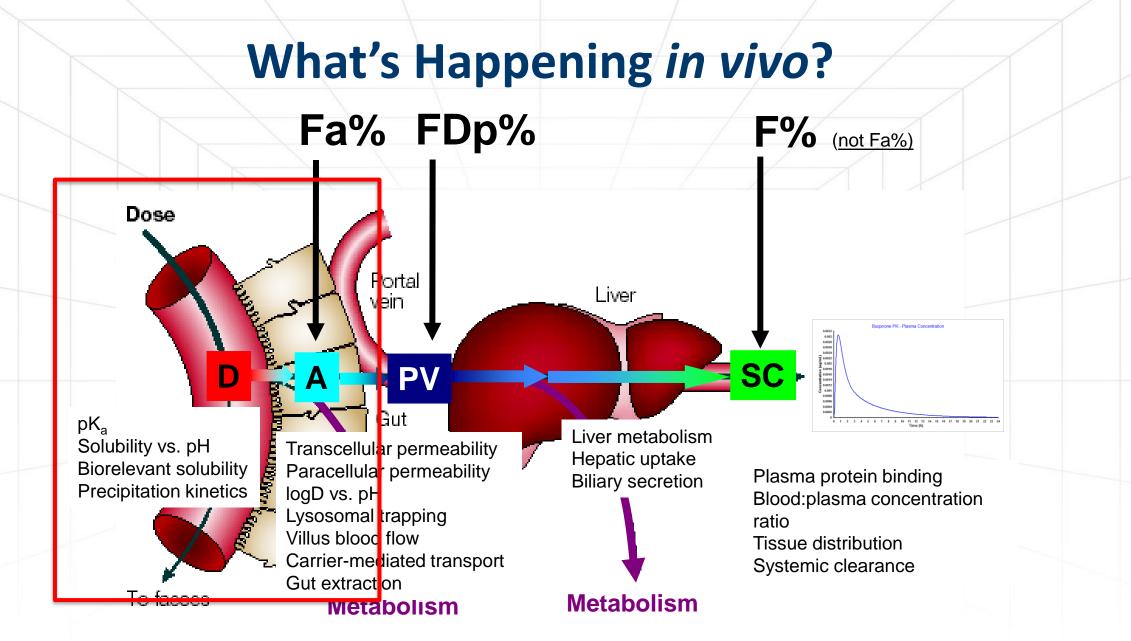
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Absorption Predictions: Current Capabilities and Knowing the Gaps

Viera Lukacova Simulations Plus, Inc. viera.lukacova@simulations-plus.com

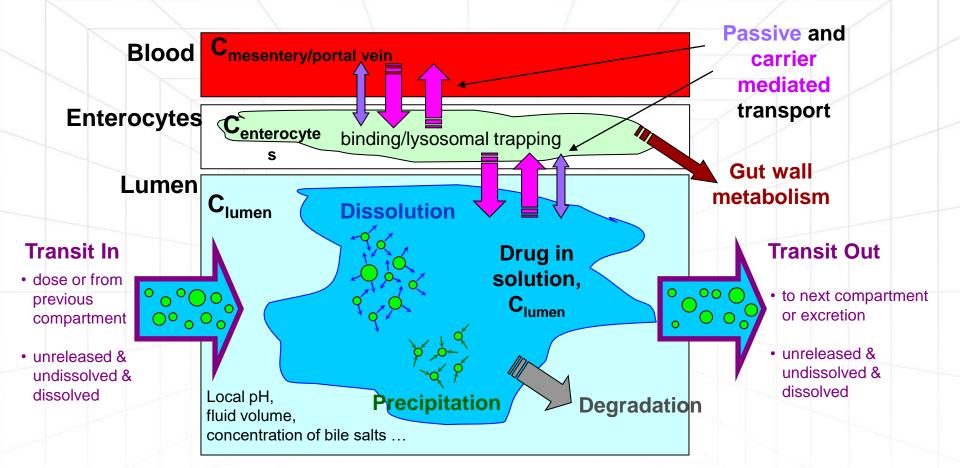
September 2022



* Modified from van de Waterbeemd, H, and Gifford, E. *ADMET In Silico Modelling: Towards Prediction Paradise?* Nat. Rev. Drug Disc. 2003, 2:192-204



Processes Involved in Oral Absorption



These phenomena:

- are happening simultaneously
- are repeated in each of the compartments of the gastrointestinal tract



Some of the Important Interactions

Dissolution–Absorption

Faster absorption can promote faster dissolution

Degradation-Absorption

Degradation reduces lumen concentration, decreasing absorption

Solubility-Absorption-Dissolution

Low solubility can limit concentration gradient and limit absorption Higher solubility increases dissolution rate

Plasma Protein Binding–Absorption

High plasma protein binding = less resistance to drug crossing the basolateral membrane

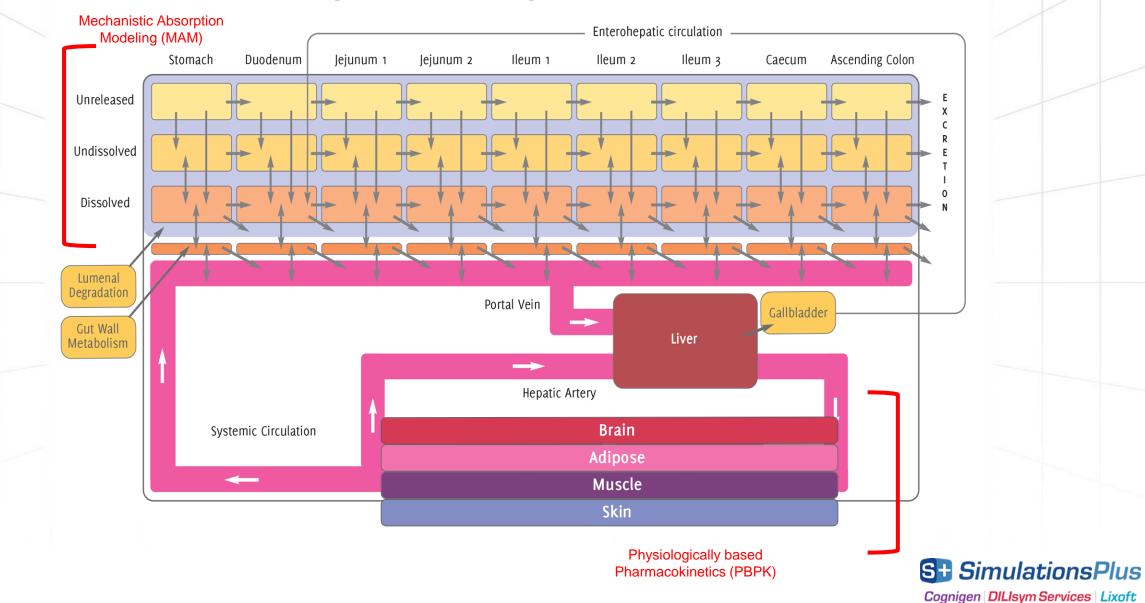
Plasma Protein Binding–Metabolism

High plasma protein binding limits metabolism

Meal (food) effects

Increased hepatic blood flow rate Gall bladder emptying Caecum emptying into colon





Advanced Compartmental Absorption and Transit Model (ACAT™)

Intestinal Physiology

in silico model needs to account for changes in conditions along gastrointestinal tract, between prandial states, and between species:

- pH
- Bile salt concentrations
- Volume of fluid
- Absorptive surface area
- Pore sizes and porosity (for paracellular absorption)
- Enzyme and transporter expression levels
- -----

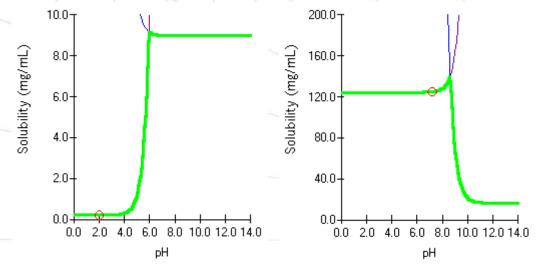






Solubility - pH

Changes in ionization result in chanes in solubility in different regions of the intestine



Changes in bile salt concentrations in different regions of the intestine may result in changes in solubility (especially for more lipophilic compounds)

$$Sol_{bile,pH} = Sol_{aq,pH} \left(1 + \frac{MWt_{H_2O}}{\rho_{H_2O}} \times SR \times C_{bile}\right)$$

Mithani, Pharm Res 1996, 13:163-167

pH and bile salt concentrations

human.

	numan.				
fasted:	Compartment Data				
i dette di	Compartment	pН	Bile Salt (mM)		
	Stomach	1.30	0.0		
	Duodenum	6.00	2.800		
	Jejunum 1	6.20	2.330		
	Jejunum 2	6.40	2.030		
	lleum 1	6.60	1.410		
	lleum 2	6.90	1.160		
	lleum 3	7.40	0.140		
	Caecum	6.40	0.0		
	Asc Colon	6.80	0.0		

rai.					
Compartment Data					
Compartment	pН	Bile Sa (mM)			
Stomach	3.90	0.0			
Duodenum	5.89	20.00			
Jejunum 1	6.13	17.29			
Jejunum 2	6.13	6.980			
lleum 1	5.93	2.820			
lleum 2	5.93	1.300			
lleum 3	5.93	1.240			
Caecum	6.58	0.0			
Asc Colon	6.23	0.0			
	Compartment Compartment Stomach Duodenum Jejunum 1 Jejunum 2 Ileum 1 Ileum 2 Ileum 3 Caecum	Compartment DCompartmentpHStomach3.90Duodenum5.89Jejunum 16.13Jejunum 26.13Ileum 15.93Ileum 35.93Caecum6.58			

rot.

Commo	utura a un t. D			
Compartment Data				
Compartment	pН	Bile Salt (mM)		
Stomach	3.00	0.0		
Duodenum	6.20	5.000		
Jejunum 1	6.20	4.050		
Jejunum 2	6.20	1.820		
lleum 1	6.40	0.610		
lleum 2	6.60	0.440		
lleum 3	6.68	0.310		
Caecum	6.75	0.0		
Asc Colon	6.45	0.0		

dog:

	Compartment Data		
fed:	Compartment	pН	Bile Salt (mM)
	Stomach	4.90	0.0
	Duodenum	5.40	14.44
	Jejunum 1	5.40	12.02
	Jejunum 2	6.00	10.46
	lleum 1	6.60	7.280
	lleum 2	6.90	5.990
	lleum 3	7.40	0.730
	Caecum	6.40	0.0
	Asc Colon	6.80	0.0

Compartment Data				
Compartment	pН	Bile Salt (mM)		
Stomach	3.20	0.0		
Duodenum	5.00	20.00		
Jejunum 1	5.10	17.29		
Jejunum 2	5.10	6.980		
lleum 1	5.94	2.820		
lleum 2	5.94	1.300		
lleum 3	5.94	1.240		
Caecum	5.90	0.0		
Asc Colon	5.51	0.0		

Compartment Data			
Compartment	pН	Bile Salt (mM)	
Stomach	5.00	0.0	
Duodenum	6.20	15.40	
Jejunum 1	6.20	12.50	
Jejunum 2	6.20	5.600	
lleum 1	6.40	1.900	
lleum 2	6.60	1.340	
lleum 3	7.05	0.950	
Caecum	7.50	0.0	
Asc Colon	6.45	0.0	

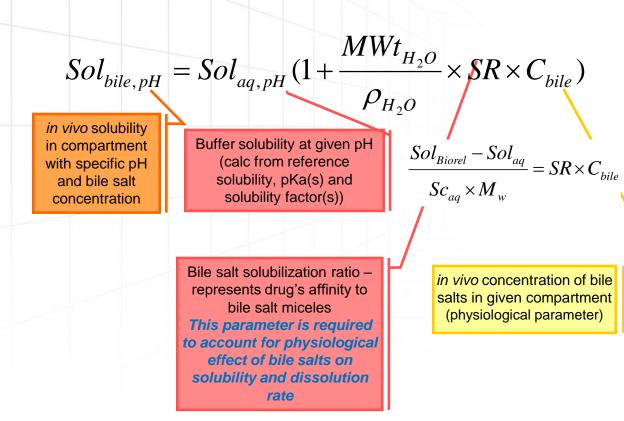


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Solubilization Ratio (SR)

in vitro value:

 measure *in vitro* solubility in media with bile salts with well defined pH and bile salt concentration (e.g. in FaSSIF or FeSSIF media) or use *in silico* estimates of these solubilities



theoretical value:

- if *in vitro* (or *in silico*) FaSSIF and FeSSIV values are not available, SR can be estimated from *logP*

 $\log SR = 2.23 + 0.61 \times \log P$

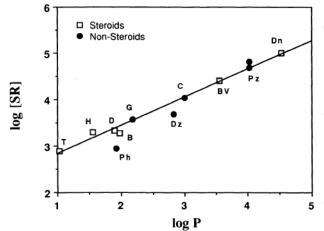


Fig. 1. Log [Solubilization Ratio] in aqueous solution taurocholate solutions as a function of log [octanol/water partition coefficient] for the non-steroidal compounds (circles; Ph = phenytoin, G = griseofulvin, Dz = diazepam, C = cyclosporin A, Pz = pentazocine), along with the prediction line based on steroid data (squares; T = triamcinolone, H = hydrocortisone, D = dexamethasone, B = betamethasone, BV = betamethasone 17-valerate, Dn = danazol); log [SR] = $2.23 + 0.60 \log [P]$.

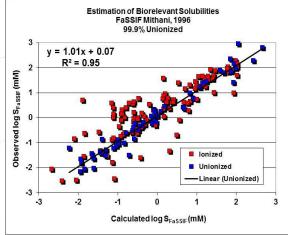
Mithani, Pharm Res 1996, 13:163-167

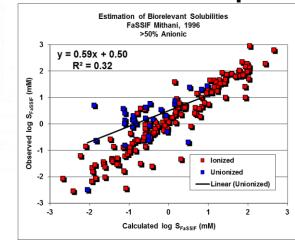


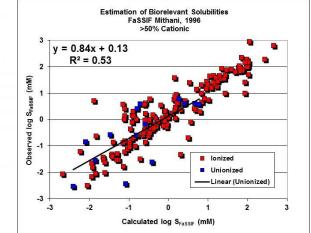
Predicting Solubilization Ratio

- The simple model for prediction of solubilization ratio from logP of the compound was evaluated using larger set of compounds (160 drug like molecules)
- The model performed well for neutral compounds, but resulted in higher errors of prediction for ionized compounds
- To estimate SR for ionized compounds, use experimental FaSSIF or FeSSIF solubilities, or *in silico* model that accounts for the ionization effects (i.e. model that was included ionized compounds in the training set)

Blue points represent different types of compounds in individual plots below Un-ionized at pH = 6.5 >50% anionic at pH = 6.5 >50% cationic at pH = 6.5







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Dissolution



Dissolution

Dissolution rate coefficient (not a constant because it changes at every time step) in intestinal lumen compartment number *i* for particle size bin *j*:

$$\frac{dM_D}{dt} = \frac{D_w}{\rho r_t T} \frac{(1+2s)}{s} (C_s - C_l) M_{u,t}$$

- D = diffusion coefficient
- C_{S} = solubility at *local* pH
- $C_{(i)}$ = lumen concentration in compartment *i*
- ρ = particle density (*density of API crystals*)
- r_i = spherical particle *radius* for particle size bin *j*
- T = diffusion layer thickness (= particle radius up to a limit)
- s = shape factor (*Length/diameter**) for spherical particles = 1

*in the original Johnson equation, s'=Length/radius and the term is $\frac{2(1+s')}{r}$



Cs

C(i)

 $T = r_{0i}$

Dissolution Models

1. Johnson (Nernst-Brunner model expanded by accounting for changing particle size due to dissolution and for non-spherical particles) $\frac{dM_D}{dt} = \frac{D_w}{\rho r_t T} \frac{(1+2s)}{s} (C_s - C_l) M_{u,t}$

Lu, Pharm Res 1993, 10:1308-1314

Wang-Flanagan (applies only to spherical particles)

$$\frac{dM_D}{dt} = \frac{3D_w}{\rho} \frac{1}{r_t} \left(\frac{1}{r_t} + \frac{1}{T}\right) (C_s - C_l) M_{u,t}$$

Wang, J Pharm Sci 1999, 88:731-738

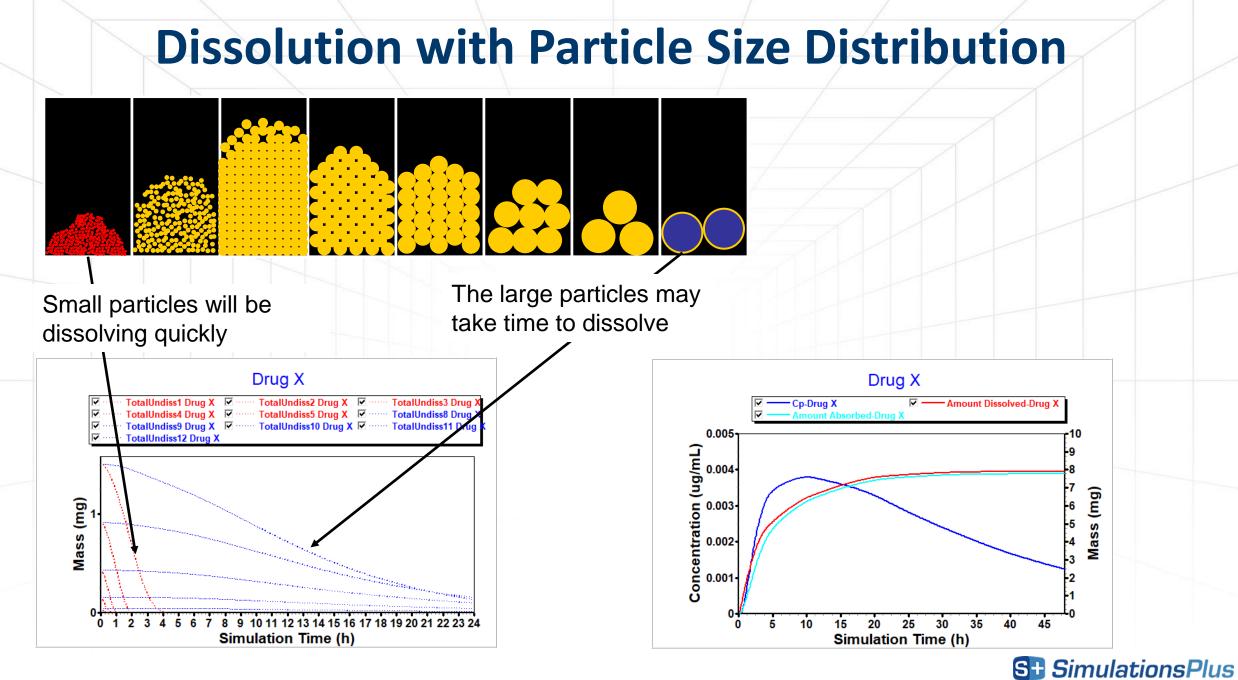
2. Z-Factor (Takano) (particle size applies ONLY through adjustment of solubility)

$$\frac{dM_D}{dt} = Z(C_s - C_l)M_{u,t}$$

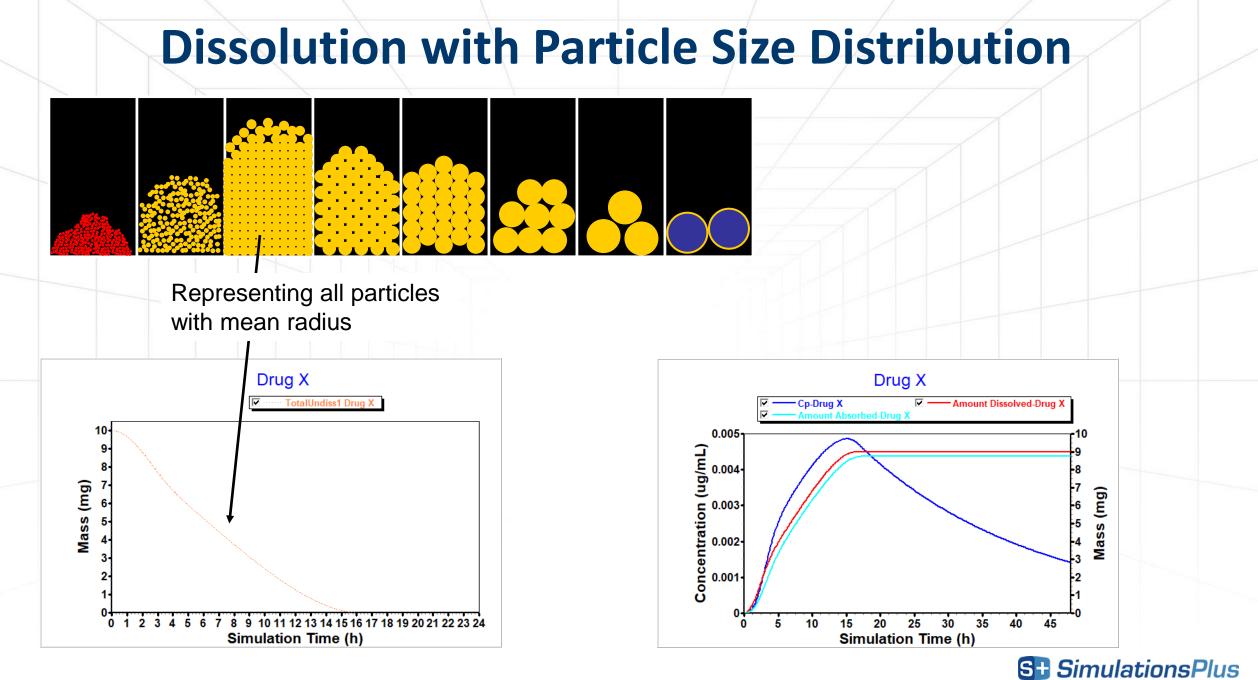
Takano, Pharm Res 2006, 23:1144-1156

z represents $\frac{3D_w}{\rho rT}$ and is determined by fitting to *in vitro* dissolution data.





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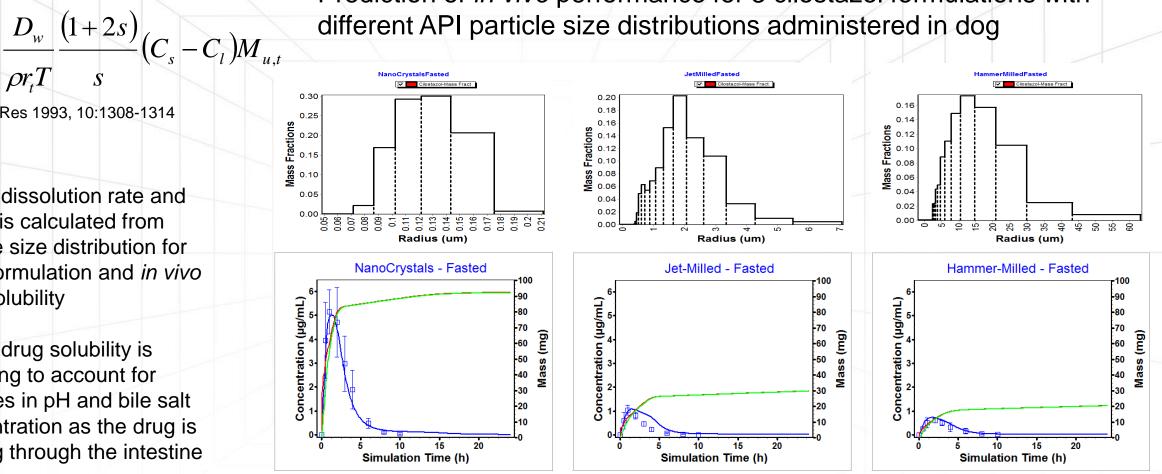
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Predicting in vivo Dissolution: Particle Size Distribution

Lu, Pharm Res 1993, 10:1308-1314

- in vivo dissolution rate and ٠ extent is calculated from particle size distribution for each formulation and in vivo drug solubility
- in vivo drug solubility is • changing to account for changes in pH and bile salt concentration as the drug is moving through the intestine

Prediction of *in vivo* performance for 3 cilostazol formulations with different API particle size distributions administered in dog



Observed data from Jinno, J Contr Rel 2006, 111: 56-64

Simulation results from GastroPlus v9.0

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 dM_D

dt

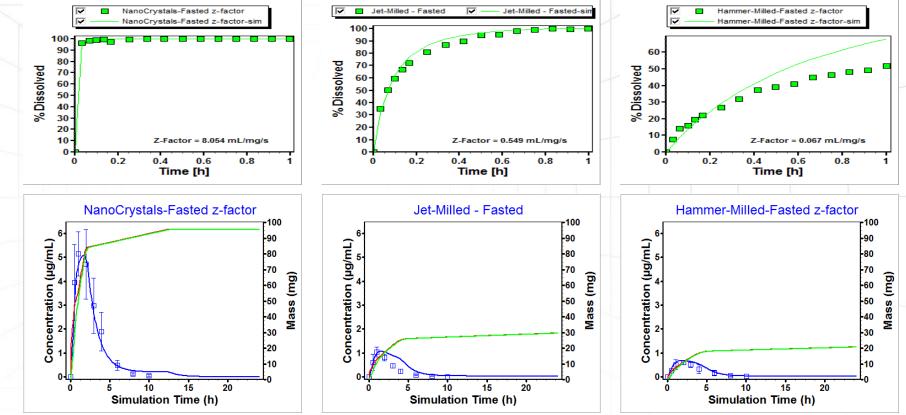
Predicting *in vivo* **Dissolution:** z-factor

Takano, Pharm Res 2006, 23:1144-1156

 $\frac{dM_D}{dt} = Z(C_s - C_l)M_{u,t}$

- *in vivo* dissolution rate and extent is calculated from zfactor fitted to *in vitro* dissolution profile for each formulation and *in vivo* drug solubility
- *in vivo* drug solubility is changing to account for changes in pH and bile salt concentration as the drug is moving through the intestine

Prediction of *in vivo* performance for 3 cilostazol formulations with different API particle size distributions administered in dog



Observed data from Jinno, J Contr Rel 2006, 111: 56-64

Simulation results from GastroPlus v9.0

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Predicting *in vivo* **Dissolution: P-PSD**

- The *in vitro* dissolution profiles showed multi-phasic behavior
- Single z-factor is not able to describe the entire dissolution profile for any of the tested batches

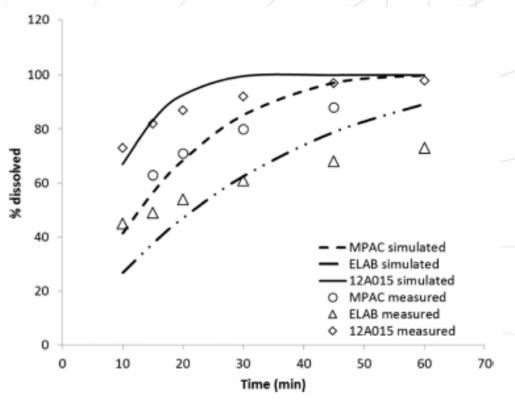


Figure 8. Z-factor fit for batches 12A015 ($Z = 1 \times 10^{-3} \text{ mL/mg/s}$), ELAB ($Z = 3.74 \times 10^{-4} \text{ mL/mg/s}$), and MPAC (Z = 5e-4 mL/mg/s).

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Predicting *in vivo* **Dissolution: P-PSD**

- Theoretical particle size distribution was fitted to one set of in vitro dissolution data
- The applicability of the fitted PSD was validated by predicting *in vitro* dissolution under different conditions
- The fitted PSD was used as an in put in *in vivo* simulation

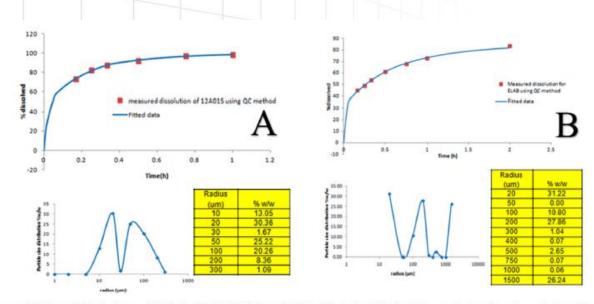
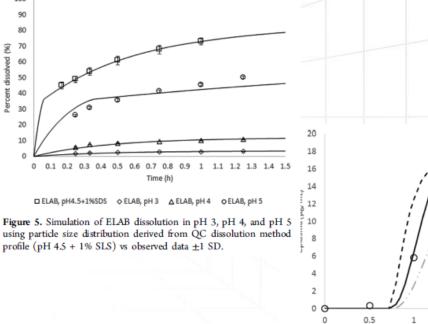


Figure 4. Fitting of dissolution profile for batch 12A015 (A) and ELAB (B) in the QC dissolution method with a theoretical particle size distribution. Note: the value presented at the 2 h time point for batch ELAB is from an infinity spin (15 min, 250 rpm).



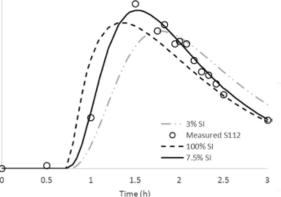


Figure 7. Simulated PK profile vs measured plasma concentrations for S112 following administration of 400 mg 12A015 tablet using Option A.



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Pepin et al. Mol Pharmaceutics 2016, 13:3256-3269

Mechanistic Models are Important for *in vitro –in vivo* Extrapolation



DDDPlus™ Simulation software for the in vitro dissolution experiment

- Extent and rate of *in vitro* dissolution for active pharmaceutical ingredient (API) and excipients:
 - Multiple particle size distributions for ingredients
 - Dynamic microclimate pH calculation
 - pH of buffers from composition of acids, bases, and salt equivalents.
 - Selection of USP and user defined experimental apparatus
 - Micelle-facilitated dissolution through addition of surfactants in medium
 - Multiple experimental phases allow for dissolution experimental design
- Differences in dosage forms:
 - IR powders, tablets, capsules, and coated beads
 - CR polymer matrix and bilayer tablet systems
 - DR coated tablets

DDDPlus(TM): DDDPlusDemo.mdb (CALLCORE\ Duk	lic\Sim \D	ע סס			
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) <u> </u>						
Formulation Name			upport File	e Informatio	n	
H Hydrocortisone Coarse Pow	▶	pport files:				
Current Record: 1; Total Records:		vdrocortisone vdrocortisone				
Oosage Form: IR: Powder	<u> </u>					
Manufacturing Properties	[I	ngredient l	nformatio	n		
Compression Force (kN): 33	5	Ingredient N	ame	Туре	Amount	
Porosity/Tortuosity: 0.52	:85	Hydrocortiso	ne	Active	150	
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Cap. Disinteg. Time (min):	_ '					
Contractor d'Instruction						
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SimulationsPlus

Precipitation



When Precipitation Plays a Role



Available online at www.sciencedirect.com

Advanced Drug Delivery Reviews 59 (2007) 568-590

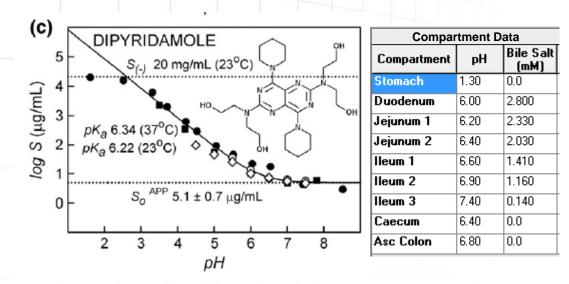


Solubility of sparingly-soluble ionizable drugs $\stackrel{\leftrightarrow}{\sim}$

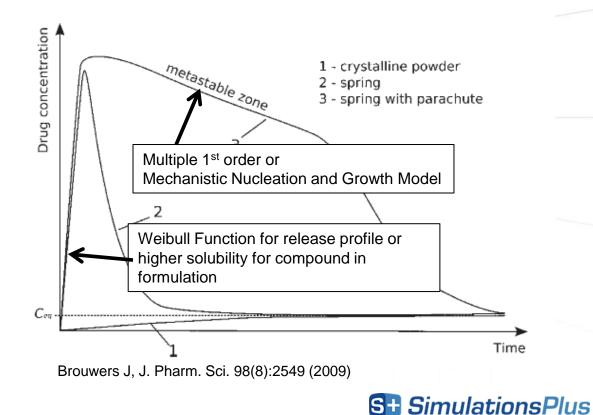
Alex Avdeef *

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Received 23 March 2007; accepted 10 May 2007 Available online 29 May 2007



Supersaturating Drug Delivery Systems
 (Salts, Cocrystals, Lipid formulations,
 SMEDDS, SNEEDS, Solid Dispersions ...)



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De Yoreo JJ, (2003): "Whether considering nucleation or growth, the reason for the transformation from solution to solid is the same, namely the free energy of the initial **bulk** solution phase is greater than the sum of the free energies of the crystalline **solid** phase plus the final solution phase (Gibbs 1876, 1878).

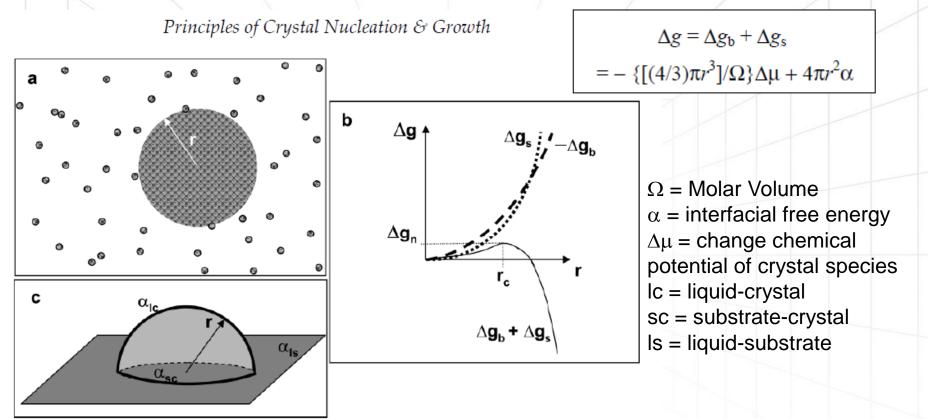


Figure 6. (a) Formation of a spherical nucleus of radius r from a solution leads to the free energy changes shown in (b). The cross-over of the bulk and surface terms combined with their opposing signs leads to a free energy barrier. (c) Heterogeneous formation of a hemispherical nucleus at a foreign substrate.

Precipitation Models

GastroPlus® First Order Precipitation: $\frac{dM_i}{dt} = \left(\frac{V_i}{T_{precip}}\right) \left(C_i - S_i\right) \qquad \begin{array}{l} M_i - \text{Mass in compartment i} \\ V_i - \text{Volume of compartment i} \\ T_p - \text{Precipitation Time} \end{array}$

Mechanistic Nucleation and Growth (different forms of this model are presented in literature):

$$\frac{dN_{nc}}{dt} = \left(\frac{R^*}{\lambda + R^*}\right) D_{mono} N_A C_{aq}^2 \left(\frac{k_B T}{\gamma}\right)^{1/2} \ln\left(\frac{C_{aq}}{S_{aq}}\right)$$
$$\times \exp\left[-ECF \times \frac{16\pi}{3} \left(\frac{\gamma}{k_B T}\right)^3 \left\{\frac{(v_m)}{\ln(C_{aq}/S_{aq})}\right\}^2\right]$$

- D_{mono} = diffusion coefficient of the monomer (3.42E-4 cm²/min)
- N_A = Avogadro's number (6.02E+23 molecs/mole)
- C_{aq} = Conc. of free monomer (moles/cm^3)
- $S_{aq} =$ Solubility at the current pH
- k_b = Boltzman's constant (1.38E-21 cJoules/Deg. K) (Note: Joule = Newton-meter)

- *T* = 310° K
- γ = Interfacial tension (Newtons/cm)
- v_m = Molecular volume = (V_m/N_A = XX cm³/molec / 6.02E+23 molec/mole)
- R^* = Critical radius (cm)
- $\lambda = \text{Effective radius from Lindfors (cm)}$
- *ECF* = exponential correction factor



How Do We Account for Excipient Effects on Nucleation?

- Lindfors, 2008
 - If the rate of association/dissociation is entirely controlled by the diffusive flow of monomers from the cluster surface to the bulk and vice versa, the net flow of monomers to a cluster is: $Q = 4\pi R D_0 (C_b S_0)$
 - Where R is the cluster radius, D_0 is the monomer diffusion coefficient, C_b is the monomer concentration in the bulk solution, and S_0 is solubility.

$$\varphi = \frac{R^*}{R^* + \lambda}$$

 $- Q = \varphi 4\pi R D_0 (C_b - S_0)$

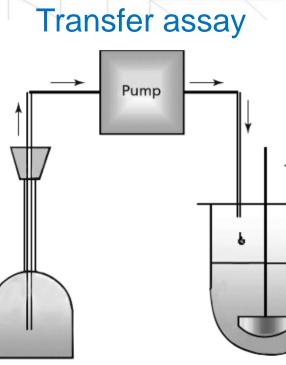
- Effective size = λ

- If $R^* >> \lambda$ the transport is controlled by diffusion,
- but if $R^* \ll \lambda$ the surface integration is the limiting process.
- $-\lambda = 0.006$ to 6.0 microns without nucleation inhibitors
- $-\lambda = 6$ to 1000 microns without PVP

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in vitro Precipitation Experiments



Donor phase



Figure 2 Experimental set-up to examine precipitation.

Figure from Kostewicz E, et al., J. Pharm. Pharmacol. 56:43 (2004)

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9			
-		-	

Membrane Dissolution

Receiver Volume (mL): Receiver Viscosity (g/(cm*s)):	20.00
Receiver Viscosity (g/(cm*s)):	
	0.01000
Receiver Diffusion Layer Thickness (um)	10.00
Donor Diffusion Layer Thickness (um):	10.00
Membrane Thickness (um):	125.00
Membrane Area (cm^2):	1.54
Membrane Porosity (fraction):	0.760

U Biphasic Dissoluton

Biphasic Dissolution

▼ Turn on Biphasic Dissolution?				
Organic Volume (mL):	40.00			
Organic Viscosity (g/(cm*s)):	0.0484			
Organic Density (g/mL):	0.830			
Aqueous Diffusion Layer (um):	50.00			
Organic Diffusion Layer (um):	50.00			
Interfacial Area (cm^2):	19.63			

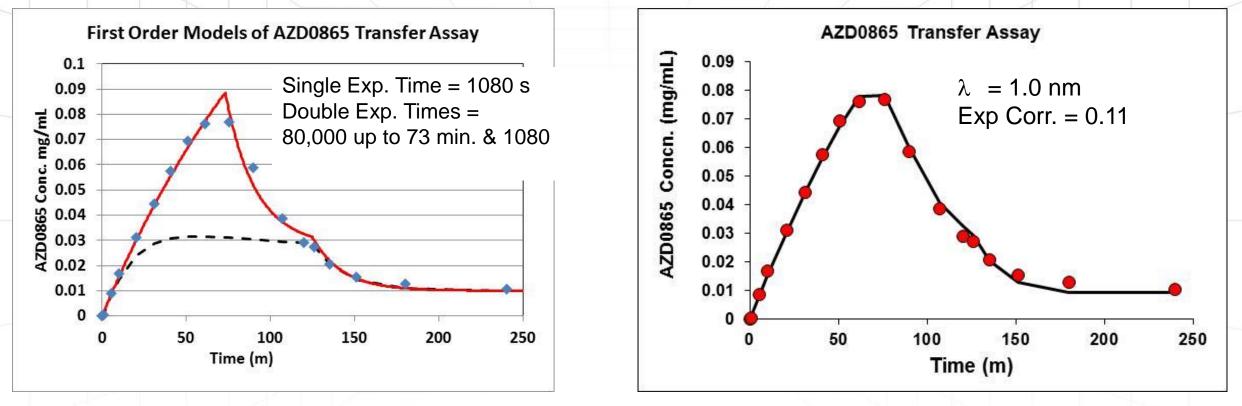
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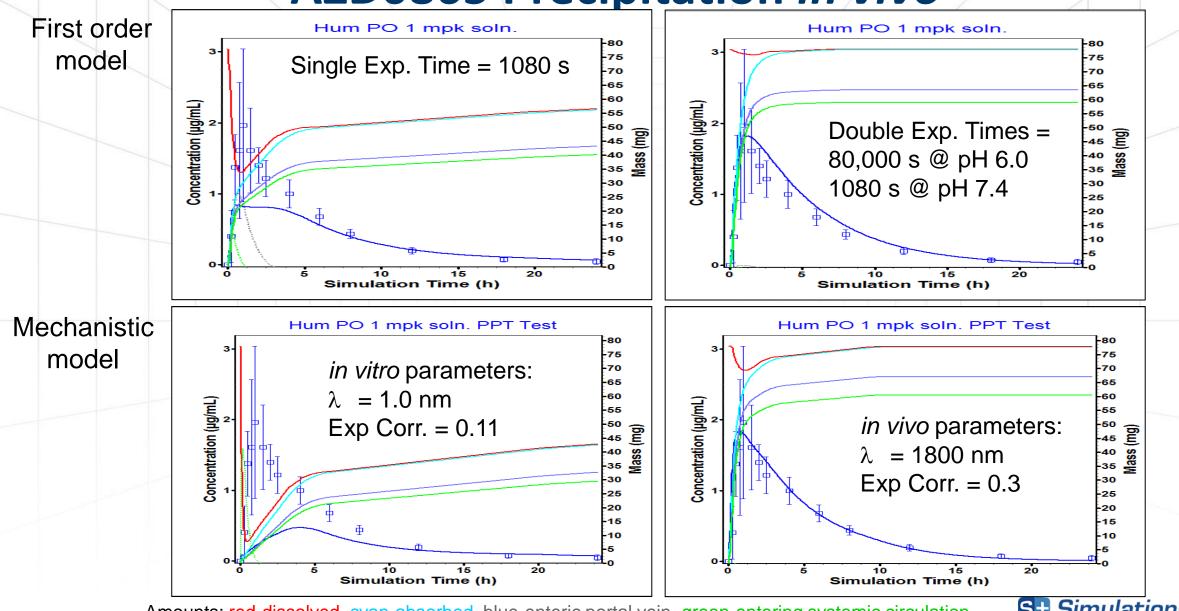
Transfer Assay



Data from Carlert S. Pharm. Res. 27:2119 (2010)



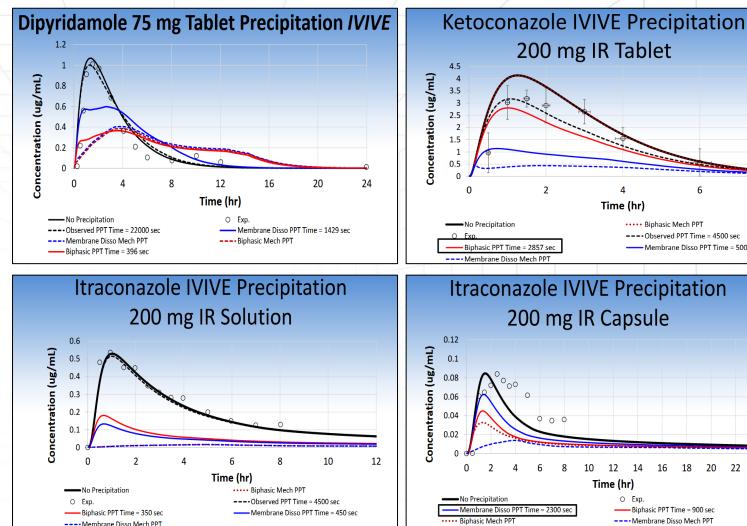
AZD0865 Precipitation in vivo



Amounts: red-dissolved, cyan-absorbed, blue-enteric portal vein, green-entering systemic circulation Dark blue line and points – plasma concentration St SimulationsPlus Cognigen | DILIsym Services | Lixoft

Biphasic and Membrane Dissolution

- Two-stage biphasic or membrane dissolution experiment were used to measure in vitro precipitation behavior
- DDDPlus[™] was used to analyze the in vitro data using first order and mechanistic precipitation models
- The in vitro parameters were used to predict the in vivo exposure
- Different in vitro experiment and model provided accurate in vivo prediction for different compound/formulation



Webinar: Mullin – Best Practices for Membrane & Biphasic In Vitro Dissolution with DDDPlus™ & GastroPlus® https://www.simulations-plus.com/resource/best-practices-for-membrane-biphasic-in-vitro-dissolution-with-dddplus-gastroplus/

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Biphasic PPT Time = 900 sec

•••• Biphasic Mech PPT

14 16 18 20

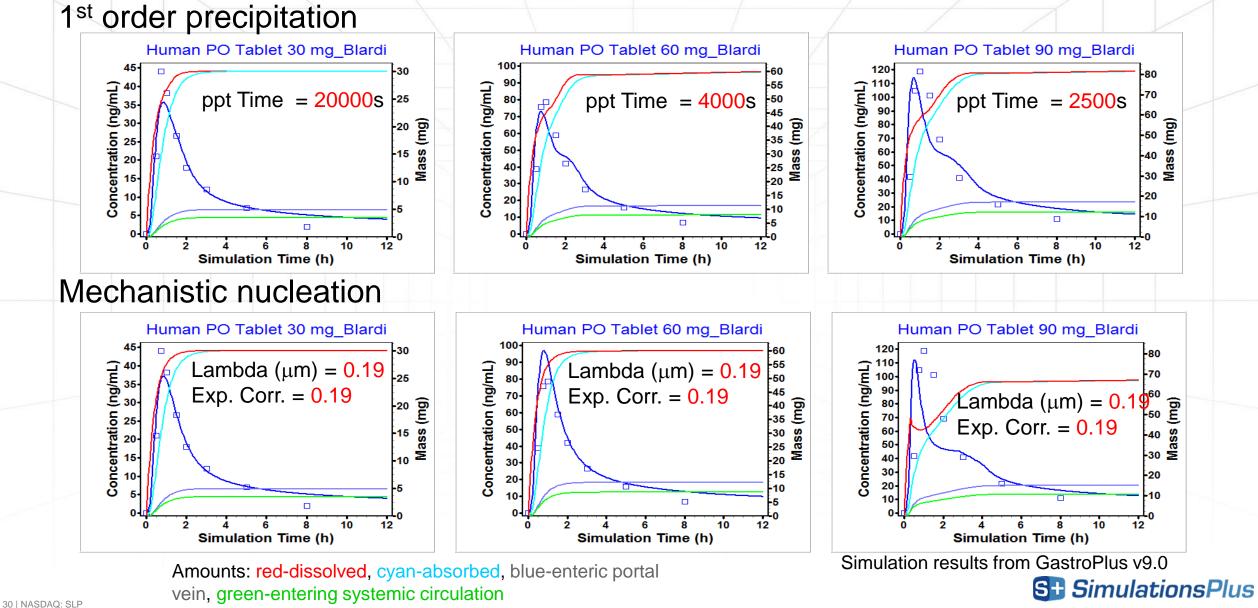
---Observed PPT Time = 4500 see

Membrane Disso PPT Time = 500 set

S⁺ Simula

22 24

Nimodipine Solid Dispersion



Dark blue line and points – plasma concentration

Data from Blardi - Clin Pharm Ther 72:556 (2002)

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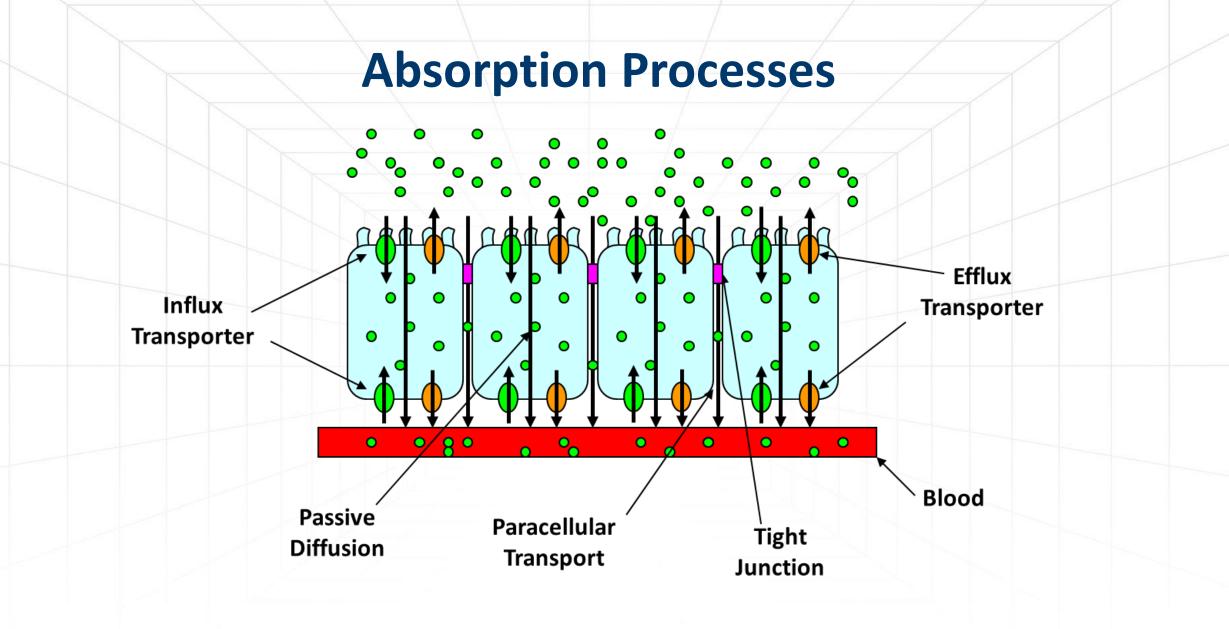
Precipitation Models

- Mechanistic nucleation and particle growth are important concepts for formulation of poorly soluble APIs.
- Testing *in vitro* is valuable for predicting the tendency to supersaturate and the relationship between supersaturation ratio and precipitation time.
- Simple 1st order methods have proven to be useful in understanding in vivo precipitation.
- Mechanistic nucleation and growth theory provides a more detailed understanding of the impact of chemistry and formulation on *in vivo* performance.
- First order precipitation time may need to change for each dose because the degree of supersaturation is different.
- One setting for Mechanistic Nucleation and Growth will explain all doses of enabled formulations.









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Effective Permeability (P_{eff}): Measurements in Human

- Measure *disappearance* of drug from donor side
- Factors affecting permeability:
 - individual subject
 variations
 - adsorption to the tubes

Peff = Q(Cin - Cout) /(2 π r L Cin) r=1.75 cm, L=10 cm Peff = 0.0091*Q(Cin - Cout) /Cin

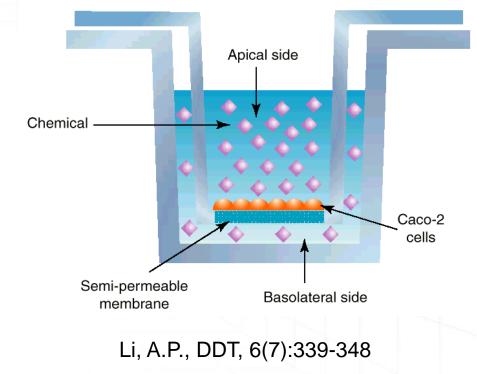
Cross-section



H. Lennernas, G.L. Amidon, et al. Capsugel Library, 1995

in vitro Permeability (P_{app}) Experiments

- Measure *appearance* of drug on receiver side
- Many factors affect in vitro permeability (P_{app}):
 - pH on each side of the membrane
 - solvents (e.g., DMSO)
 - amount of protein on receiver side
 - concentration in donor side
 - shaking rate
 - nonspecific binding to plasticware

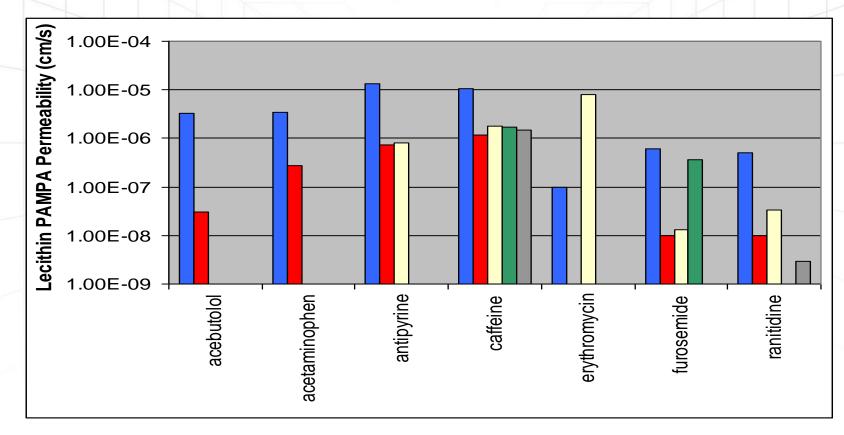




Variability of in vitro Permeability

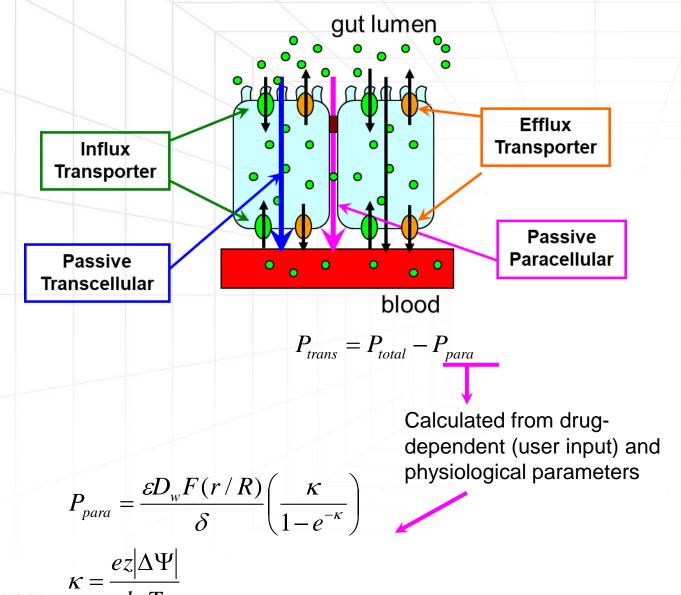
PAMPA Permeability values for selected drugs collected from literature All data are for Lecithin solution in dodecane and pH=7.4

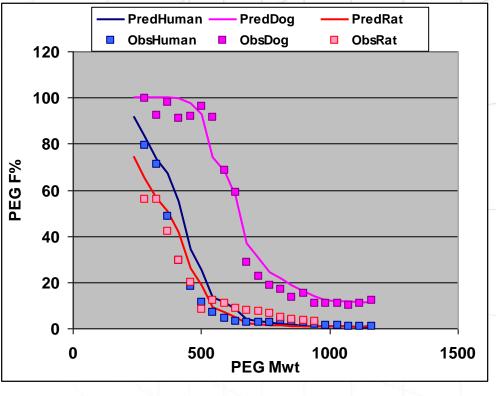
Blue – Zhu Ch. EurJMedChem 2002, 37:399; Red – Ruell J.A. pION; Yellow – Kerns E.H. JPharmSci 2004, 93:1440; Green – Avdeef A. EurJPharmSci 2001, 14:271; Grey – Du C. pION



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Paracellular Permeability in Absorption





Data from He-JPharmSci 1998, 87: 626-633

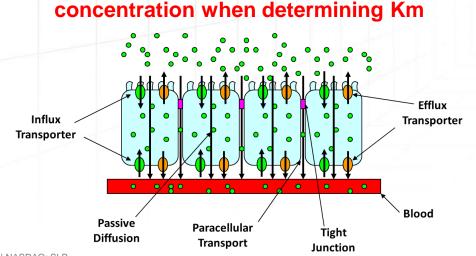
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Carrier-Mediated Absorption

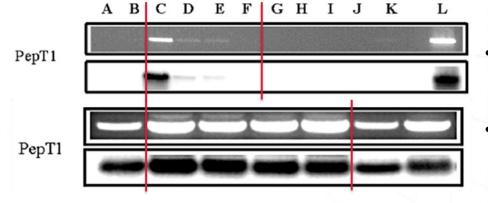
- Transporter expressions vary along the intestine
- Transporter expression patterns vary among species and differ from *in vitro*
- Considerable variability in reported expression patterns from different sources
- New measurements are being reported

Be mindful of the effective



Compartment	PepT1 ^a	PepT1 ^b	PepT1 ^c	HPT1 ^b	P-gp ^a	$P-gp^d$	OCTN1 ^c	LAT2 ^e	OATP 1A2
Duodenum	0.72	8.42	1.02	1.06	0.16	0.70	0.42	0.74	0.07
Jejunum1	0.92	1.48	1.01	1.17	1.23	0.84	0.71	4.57	0.07
Jejunum2	0.92	1.48	1.01	1.17	1.23	0.94	0.71	4.57	0.07
Ileum1	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00
Ileum2	1.00	1.00	1.00	1.00	1.00	1.09	1.00	1.00	1.00
Ileum3	1.00	1.00	1.00	1.00	1.00	1.07	1.00	1.00	1.00
Caecum	0.02	0	0.04	1.36	0.36	1.3	0.34	0	0.13
Asc. Colon	0.02	0	0.04	1.70	0.36	1.3	0.34	0	0.17

Bolger MB, AAPS Journal, 11(2):353 (2009)



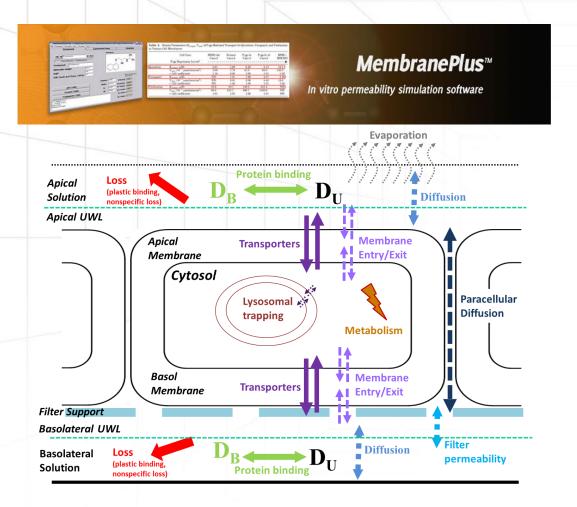
Herrera-Ruiz AAPS Pharmsci 2001; 3 (1) article 9 (http://www.aapspharmaceutica.org)

- RT-PCR analysis of human (top) and rat (bottom) PepT1
- Red lines mark small intestine (duodenumileocaecal junction)

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Mechanistic Models are Important for *in vitro –in vivo* Extrapolation

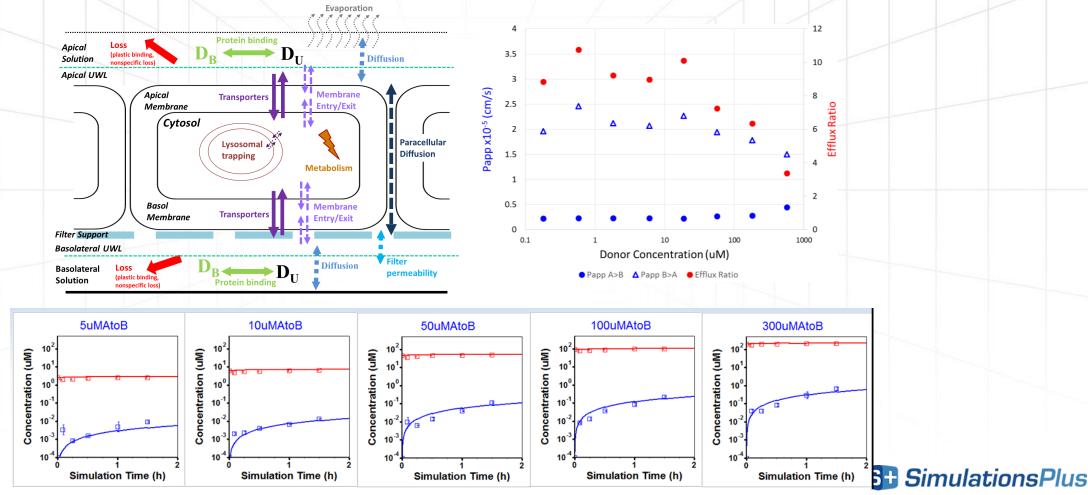


- Predict *in vitro* Papp and analyze measured data to unlock important information related to absorption
- Number of different processes affecting apparent *in vitro* permeability can be included in the simulation:
 - Passive transcellular diffusion
 - Passive paracellular diffusion
 - Carrier mediated influx and/or efflux
 - Metabolism in the cells
 - Binding to albumin
 - Accumulation in cell membranes or intracellular compartments
 - Experimental conditions (e.g., shaking rate, pH)

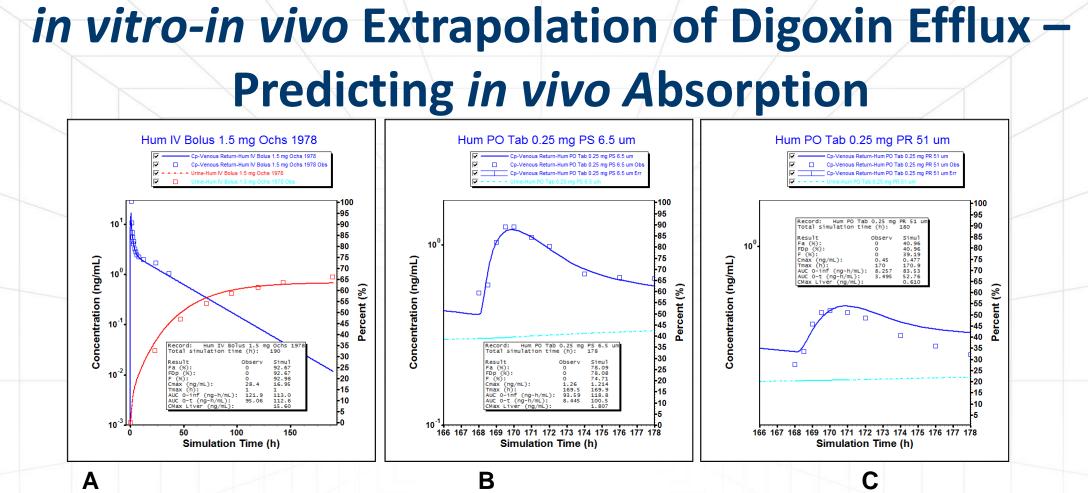


in vitro-in vivo Extrapolation of Digoxin Efflux – Determining Intracellular Unbound Km

The intracellular unbound P-gp K_m for digoxin was found to be 95.3 µM by fitting B->A Papp with MembranePlus[™] across experiments run at eight different concentrations and validated in a separate experiment using kinetic data at five concentrations



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A: Observed (symbols) vs. predicted plasma conc. (blue) and urinary excretion (red) of digoxin (Ochs, 1978).

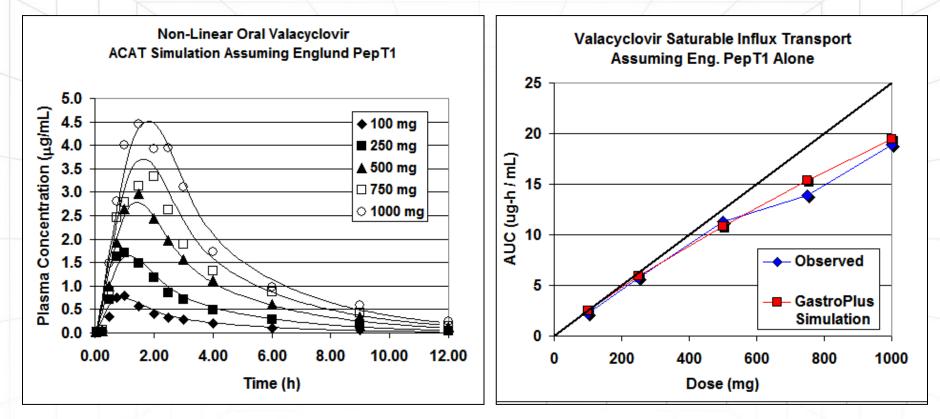
B: Observed (symbols) vs. predicted plasma conc. (blue) of digoxin for a PO formulation with 6.5 μm radius particle size (Jounela, 1975).

C: Observed (symbols) vs. predicted plasma conc. (blue) of digoxin for a PO formulation with 51 μ m radius particle size (Jounela, 1975).

All simulations are using the fitted intracellular unbound P-gp Km value of 95.3 mM



GastroPlus Simulation of Nonlinear Dose Dependence for Influx Transport of Valacyclovir



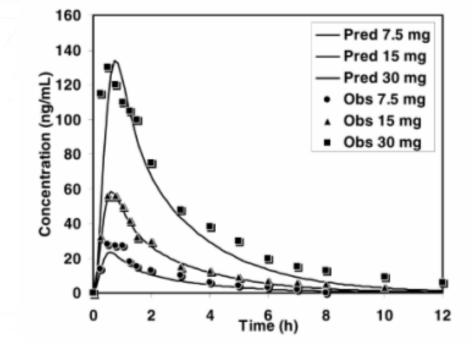
Bolger MB, et al. AAPS Journal 11(2):353 (2009) GastroPlus results were first reported in Feb. 2003 at AAPS Drug Transport Workshop, Peachtree City, GA



Remember the Impact of Intestinal Metabolism

- Enzyme expressions vary along the intestine
- Enzyme expression patterns vary among species
- *in vitro-in vivo* extrapolation is established better than for transporters
- Compound elimination in the intestine affects the concentration and impacts the other processes

GastroPlus simulations of nonlinear dose dependence for midazolam using *in vitro* K_m and V_{max} and *iv* PK (Agoram et al., 2001)



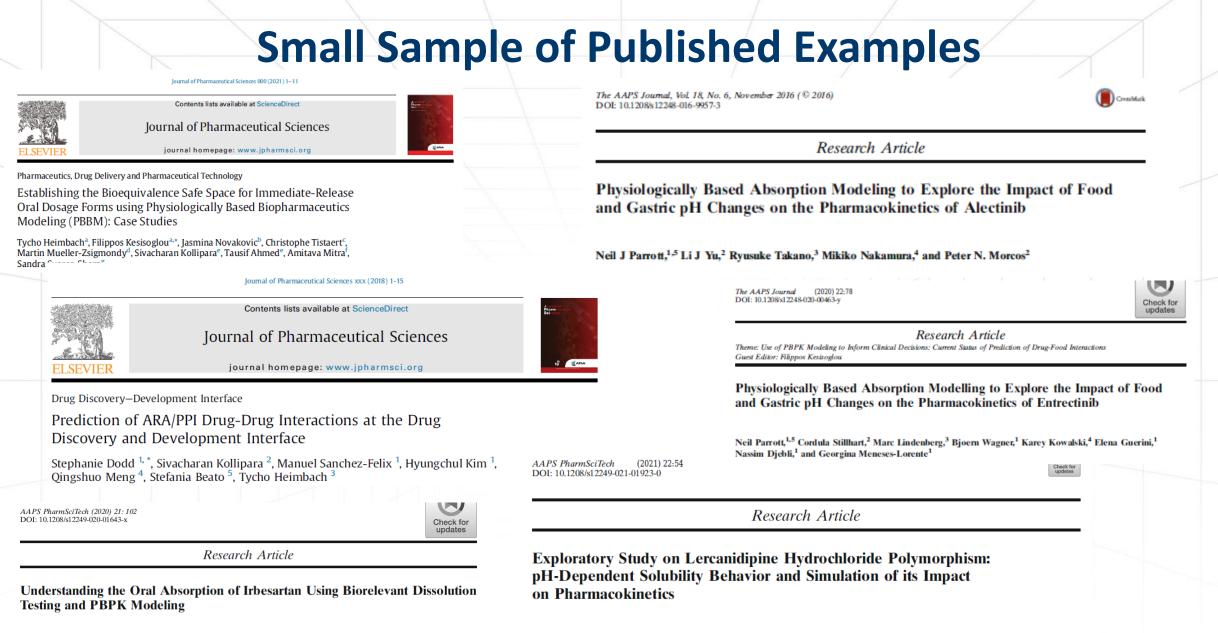
DoseCmaxAUCCmaxAUCFa%FDP%Fb%7.50.028690.02165994524150.0561540.052158995529300.134530.120369996434		Experiment	al	GastroPlus	Compartm	ental Simul	ated	
150.0561540.052158995529	Dose	Cmax	AUC	Cmax	AUC	Fa%	FDP%	Fb%
	7.5	0.028	69	0.021	65	99	45	24
30 0.13 453 0.120 369 99 64 34	15	0.056	154	0.052	158	99	55	29
	30	0.13	453	0.120	369	99	64	34

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Summary

- Number of different processes contribute to net drug absorption
- Different *in vitro* assays are available to determine input parameters for individual processes
- Mechanistic simulations of *in vitro* assays are an important tool to 'deconvolute' parameter values and aid in *in vitro* – *in vivo* extrapolation
 - Accuracy of *in vitro in vivo* extrapolation varies between the processes
- To predict the overall *in vivo* absorption, the interplay of all relevant processes needs to be considered





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Navpreet Kaur,¹ Poonam Singh Thakur,² Ganesh Shete,³ Rahul Gangwal,⁴

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Small Sample of Published Examples

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ARTICLE

Application of physiologically based biopharmaceutics modeling to understand the impact of dissolution differences on in vivo performance of immediate release products: The case of bisoprolol

Joyce S. Macwan¹ | Grace Fraczkiewicz¹ | Mauro Bertolino² | Phillip Krüger³ Sheila-Annie Peters²

AAPS J (2022) 24:17 https://doi.org/10.1208/s12248-021-00663-0

Research Article

Theme: Integrating In Vitro Systems and Physiologically-Based Pharmacokinetics Modeling to Optimize Drug Product Development Guest Editors: Rodrigo Cristofoletti and Lawrence Yu

Dissolution Challenges Associated with the Surface pH of Drug Particles: Integration into Mechanistic Oral Absorption Modeling

Bart Hens^{1,6}, Nidhi Seegobin^{1,2}, Marival Bermejo³, Yasuhiro Tsume⁴, Nicola Clear¹, Mark McAllister¹, Gregory E. Amidon⁵, Gordon L. Amidon^{5,6}

European Journal of Pharmaceutical Sciences 47 (2012) 375-386



Contents lists available at SciVerse ScienceDirect European Journal of Pharmaceutical Sciences

journal homepage: www.elsevier.com/locate/ejps

Application of PBPK modeling to predict human intestinal metabolism of CYP3A substrates – An evaluation and case study using GastroPlus™

Aki T. Heikkinen, Guillaume Baneyx, Antonello Caruso, Neil Parrott ' F. Hoffmann-La Roche AG, pRED, Pharma Research & Early Development, Non-Clinical Safety, Basel, Switzerland



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BCS Class IV Oral Drugs and Absorption Windows: Regional-Dependent Intestinal Permeability of Furosemide

Milica Markovic 10, Moran Zur 1, Inna Ragatsky 1, Sandra Cvijić 20 and Arik Dahan 1,*0

European Journal of Pharmaceutical Sciences 155 (2020) 105552

- Contents lists available at ScienceDirect European Journal of Pharmaceutical Sciences
 - journal homepage: www.elsevier.com/locate/eips

A combined in vitro in-silico approach to predict the oral bioavailability of borderline BCS Class II/IV weak base albendazole and its main metabolite albendazole sulfoxide

Maximo Pettarin^a, Michael B. Bolger^b, Maja Chronowska^a, Edmund S. Kostewicz^{a,}



pharmaceutics

PBPK Modeling as a Tool for Predicting and Understanding Intestinal Metabolism of Uridine 5'-Diphospho-glucuronosyltransferase Substrates

Micaela B. Reddy ^{1,*}, Michael B. Bolger², Grace Fraczkiewicz², Laurence Del Frari³, Laibin Luo⁴, Viera Lukacova², Amitava Mitra⁵, Joyce S. Macwan², Jim M. Mullin², Neil Parrott⁶ and Aki T. Heikkinen⁷

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Theme: Towards Integrated AD ME Prediction: Past, Present, and Future Directions Guest Editors: Lawrence X. Yu, Steven C. Sutton, and Michael B. Bolger

Simulations of the Nonlinear Dose Dependence for Substrates of Influx and Efflux Transporters in the Human Intestine

Michael B. Bolger,^{1,2,3} Viera Lukacova,¹ and Walter S. Woltosz

The AAPS Journal (2020) 22:134 DOI: 10.1208/s12248-020-00511-7

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Research Article

Theme: Use of PBPK Modeling to Inform Clinical Decisions: Current Status of Prediction of Drug-Food Interactions Guest Editor: Filippos Kesisoglou

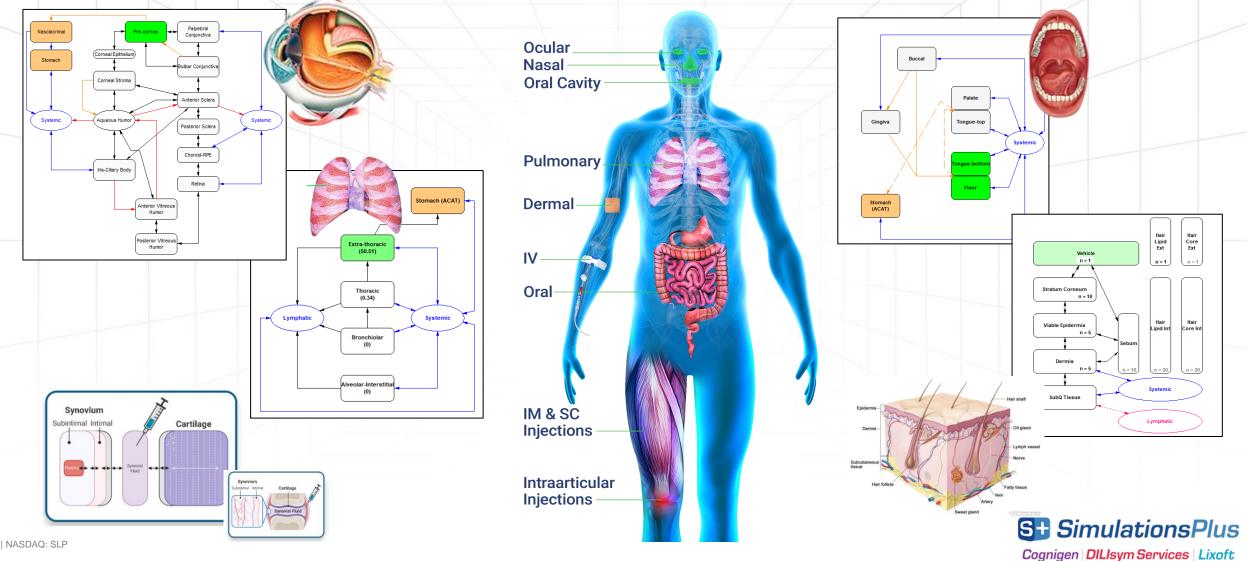
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MDPI

Physiologically Based Pharmacokinetic Modeling of Oral Absorption, pH, and Food Effect in Healthy Volunteers to Drive Alpelisib Formulation Selection

Monika Gajewska,¹ Lars Blumenstein,¹ Alexandros Kourentas,² Martin Mueller-Zsigmondy,² Sebastien Lorenzo,³ Angela Sinn,⁴ Maria Velinova,⁵ and Tycho Heimbach⁶

Similar Considerations Apply to Other Routes of Administration...



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